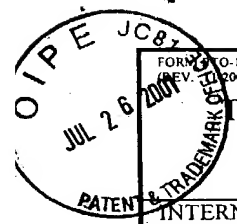


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FORM PTO-1390 (REV. 12-2000) U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE		ATTORNEY'S DOCKET NUMBER 266/035 U.S. APPLICATION NO. (If known, see 37 CFR 1.5) Not Yet Assigned 097890335
TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371		
INTERNATIONAL APPLICATION NO. PCT/EP00/00597	INTERNATIONAL FILING DATE 26 January 2000	PRIORITY DATE CLAIMED 27 January 1999
TITLE OF INVENTION Noninvasive Vaccination Through the Skin		
APPLICANT(S) FOR DO/EO/US Gregor Cevc et al		
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:		
1. <input checked="" type="checkbox"/> This is a FIRST submission of items concerning a filing under 35 U.S.C. 371. 2. <input type="checkbox"/> This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371. 3. <input checked="" type="checkbox"/> This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (21) indicated below. 4. <input checked="" type="checkbox"/> The US has been elected by the expiration of 19 months from the priority date (Article 31). 5. <input checked="" type="checkbox"/> A copy of the International Application as filed (35 U.S.C. 371(c)(2)) a. <input checked="" type="checkbox"/> is attached hereto (required only if not communicated by the International Bureau). b. <input type="checkbox"/> has been communicated by the International Bureau. c. <input type="checkbox"/> is not required, as the application was filed in the United States Receiving Office (RO/US). 6. <input type="checkbox"/> An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)). a. <input type="checkbox"/> is attached hereto. b. <input type="checkbox"/> has been previously submitted under 35 U.S.C. 154(d)(4). 7. <input checked="" type="checkbox"/> Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)) a. <input type="checkbox"/> are attached hereto (required only if not communicated by the International Bureau). b. <input type="checkbox"/> have been communicated by the International Bureau. c. <input type="checkbox"/> have not been made; however, the time limit for making such amendments has NOT expired. d. <input checked="" type="checkbox"/> have not been made and will not be made. 8. <input type="checkbox"/> An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371 (c)(3)). 9. <input type="checkbox"/> An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)). 10. <input type="checkbox"/> An English language translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)). Items 11 to 20 below concern document(s) or information included: 11. <input type="checkbox"/> An Information Disclosure Statement under 37 CFR 1.97 and 1.98. 12. <input type="checkbox"/> An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included. 13. <input type="checkbox"/> A FIRST preliminary amendment. 14. <input type="checkbox"/> A SECOND or SUBSEQUENT preliminary amendment. 15. <input type="checkbox"/> A substitute specification. 16. <input type="checkbox"/> A change of power of attorney and/or address letter. 17. <input type="checkbox"/> A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825. 18. <input type="checkbox"/> A second copy of the published international application under 35 U.S.C. 154(d)(4). 19. <input type="checkbox"/> A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4). 20. <input checked="" type="checkbox"/> Other items or information: PCT Cover Page of Published Int'l Application PCT Notification of Transmittal of IPER PCT Notification of International Search Report PCT Request PCT Written Opinion Return Postcard		

U.S. APPLICATION NO. (PCT/EP) 09/890335		INTERNATIONAL APPLICATION NO. PCT/EP00/00597		ATTORNEY'S DOCKET NUMBER 266/035	
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21. <input type="checkbox"/> The following fees are submitted: BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)): Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO. \$1000.00 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$860.00 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$710.00 International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$690.00 International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) \$100.00 ENTER APPROPRIATE BASIC FEE AMOUNT =				CALCULATIONS PTO USE ONLY	
				\$ 860.00	
Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).				\$ 130.00	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	\$	
Total claims	- 20 =		x \$18.00	\$	
Independent claims	- 3 =		x \$80.00	\$	
MULTIPLE DEPENDENT CLAIM(S) (if applicable)				+	\$270.00
TOTAL OF ABOVE CALCULATIONS =					\$ 990.00
<input type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.				+	\$
SUBTOTAL =					\$ 990.00
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).					\$
TOTAL NATIONAL FEE =					\$
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +					\$
TOTAL FEES ENCLOSED =					\$
				Amount to be refunded:	\$
				charged:	\$

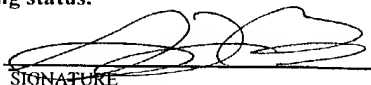
a. ☐ A check in the amount of \$ _____ to cover the above fees is enclosed.

b. ☒ Please charge my Deposit Account No. 12-2475 in the amount of \$ 990.00 to cover the above fees.
 A duplicate copy of this sheet is enclosed.

c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any
 overpayment to Deposit Account No. 12-2475. A duplicate copy of this sheet is enclosed.

d. ☐ Fees are to be charged to a credit card. **WARNING:** Information on this form may become public. **Credit card
 information should not be included on this form.** Provide credit card information and authorization on PTO-2038.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR
 1.137 (a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO: Suzanne L. Biggs Lyon & Lyon 633 West Fifth Street, Suite 4700 Los Angeles, CA 90071	 SIGNATURE Suzanne L. Biggs NAME <u>30,158</u> REGISTRATION NUMBER
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14/PRTS

IDEA AG
Our Ref.: C 2260 PCT



Noninvasive vaccination through the skin

The present invention relates to novel vaccines for the non-invasive, transcutaneous administration of antigens associated with ultradeformable carriers, for the purpose of prophylactic or therapeutic vaccination. The vaccines comprise (a) a transdermal carrier which is a penetrant, suspended or dispersed in an aqueous solvent in the form of a minute fluid droplet surrounded by a membrane-like coating of one or several layers of at least two different substances or two different forms of a substance with the tendency to aggregate, said substances or forms of a substance differing by at least the factor of 10 in solubility in a preferably aqueous, liquid medium, such that the average diameter of homo-aggregates of the more soluble substance or form of the substance or the average diameter of the hetero-aggregates consisting of both said substances or forms of said substance is smaller than the average diameter of homo-aggregates of the less soluble substance or form of the substance, and/or wherein the more soluble component tends to solubilise the penetrating droplet and wherein the content of such component amounts to up to 99 mol-% of the concentration required to solubilise the droplet or else corresponds to up to 99 mol-% of the saturating concentration in the un-solubilised droplet, whichever is higher, and/or wherein the elastic deformation energy of the droplet surrounding the membrane-like coating is at least 5x lower, more preferably is at least 10x lower and ideally is more than 10x lower than that of the red blood cells or of the phospholipid bilayers with fluid aliphatic chains, (b) a compound which specifically releases or specifically induces cytokine or anti-cytokine activity or exerts such an activity itself, and (c) an antigen or an allergen. The invention further relates to methods for corresponding therapeutic or prophylactic vaccination of mammals.

Several documents are cited throughout the text of this specification. Each of the documents cited herein (including any manufacturer's specifications, instructions, etc.) are hereby incorporated by reference; however, there is no admission that any document cited is indeed prior art of the present invention. Further incorporated by reference is the complete disclosure content of the co-pending application filed in the name of IDEA AG and bearing the title "Transnasal transport/immunization with highly adaptable carriers".

Skin is the best accessible, but also the most difficult, entry into the body, due to the presence of the stratum corneum. This horny layer of the skin is an evolutionary optimized barrier which resembles the blood vessel wall, in that it comprises flaccid, tightly packed and laterally overlapping cells, whereby the basic cellular-tile motif in the stratum corneum is repeated 20-30 times. The intercellular contacts in the skin, moreover, are sealed with the densely packed and well organized blend of lipids. The stratum corneum therefore not only protects the organism from infections but also precludes an efficient uptake of antigens through the skin. This fact, which is advantageous from the point of allergy, prevented successful immunization or vaccination through the intact skin to date.

The largest drugs on the market in any transdermal delivery device are smaller than 350 Da (Cevc, G. Drug delivery across the skin, Exp. Opin. Invest. Drugs (1997) 6: 1887-1937), as only such molecules can cross the tiny, self-sealing pores in the skin. The latter normally are less than 1 nm wide, when hydrophilic, or narrower, when hydrophobic. Organisms such as helminths therefore gain access into the body by penetrating the skin by using their biochemical machinery for the purpose of 'drilling holes' through the organ. Naturally occurring micro-lesions and shunts (such as pilosebaceous units) are available in the skin as well. However, they only cover up 0.1% to 0.5% of the skin surface and, consequently, do not contribute much to transcutaneous transport the fact notwithstanding that bacteria typically exploit such a route for a topical infection (Strange, P., Skov, L., Lisby, S., Nielsen, P. L., Baadsgard, O. Staphylococcal enterotoxin B applied on intact normal and intact atopic skin induces dermatoma. Arch. Dermatol. (1996) 132: 27-33.)

Only a few haptens exposed on the skin elicit a cutaneous immune response. This confirms that only sufficiently small molecules from a large load of the topically deposited haptens can find their way into the skin in an appreciable quantity. Such haptens then first irritate the organ and finally may cause hypersensitivity and contact dermatitis (Kondo, S., Sauder, D.N. Epidermal cytokines in allergic contact dermatitis. J. Am. Acad. Dermatol. (1995) 33: 786-800; Nasir, A., Gaspari, A. A. Contact dermatitis. Clinical perspectives and basic mechanisms. Clin. Rev. Allergy and Immunol. (1996) 14: 151-184). The problem is most serious with the low molecular

weight chemicals or with the pharmaceuticals combined with skin irritants, such as skin permeation enhancers (Cevc, 1997, *op. cit.*). Large molecules seldom are allergenic on the skin, owing to their limited ability to cross the barrier. A Th2 response to a highly immunogenic ovalbumin (Wang, L.-F., Lin, J.-Y., Hsieh, K.-H., Lin, R.-H. Epicutaneous exposure of protein antigen induces a predominant Th2-like response with IgE production in mice. *J. Immunol.* (1996) 156: 4079-4082.) or to Cholera toxin (Glenn, G. M., Rao, M. Matyas, (1998) 391: 851; Glenn, G.M., Scharton-Karsten T, Vasell R, Mallet C.P., Hale T.L. and Alving C.R. Transcutaneous Immunization with Cholera toxin Protects Mice Against Lethal Mucosal Toxin Challenge. *J. Immunol* (1998) 161: 3211-3214.) was possible only after an epicutaneous exposure to a large amount of such proteins and was fairly weak. Moreover, the stratum corneum elimination from the skin was a prerequisite for producing detectable quantities of the specific antibodies against adenoviruses encoding the human carcinoembryonic antigen or human GM-CSF gene in 96% or 43%, respectively, of epicutaneously treated C57BL/6 mice (Deng, H., Qun, L., Khavari, P. A. Sustainable cutaneous gene delivery. *Nature Biotechnology* (1997) 15: 1388-1390.).

No protection against the above mentioned or other epicutaneously employed antigens was reported to date. Antibodies against diphtheria or tetanus toxoid, and bovine serum albumin, which were generated by applying the antigens on the skin of BALB/c mice in combination with cholera toxin (Glenn et al., 1998, *op. cit.*) resulted in a very weak immune response without the adjuvant. Even after the inclusion of Cholera toxin (CT), the average specific antibody titre for diphtheria and tetanus antigens was around 50x and between 70x and 4000x (depending on the inclusion of individual data points), respectively, below that elicited by cholera toxin per se (Glenn et al., 1998, *op. cit.*). The corresponding absolute respective titre values were 14 ± 17 and 8 ± 16 ; the anti-BSA titre was approximately 11 ± 11 (average value \pm standard deviation as calculated from the published figures). No therapeutic or prophylactic effect was demonstrated for these low titres, which shows that the path towards simple non-invasive vaccination is not at all straightforward. The more recent paper published by the same group (Glenn et al., 1998b) demonstrated protection against CT after transnasal challenge which does not allow any conclusion with regard to protection obtainable by transdermal vaccination.

Earlier publications report on the deliverance of proteins across the skin, several orders of magnitude more efficient than in the above mentioned study, as judged by the titres, exploiting mechanosensitive and hydrosensitive, self-regulating carriers (Transfersomes) (for a review, see Cevc, 1997, *op. cit.*). For potent antigens this induced antibody titres that were comparable with those elicited by subcutaneous protein injections: in the case of BSA, the absolute titre of IgG was around 200 in either case (Paul, A., Cevc, G. Non-invasive administration of protein antigens. Epicutaneous immunization with the bovine serum albumin. *Vaccine Res.* (1995) 4: 145-164) and for gap junction protein titres between 15.000 and 100.000 were measured (Paul, A., Cevc, G., Bachhawat, B. K. Transdermal immunization with large proteins by means of ultradeformable drug carriers. *Eur. J. Immunol.* (1995) 25: 3521-3524; Paul, A., Cevc, G., Bachhawat, B. K. Transdermal immunization with an integral membrane component, gap junction protein, by means of ultradeformable drug carriers, Transfersomes. *Vaccine* (1997) 16: 188-195.). Yet, the generation of a protective immune response was not demonstrated in either of these publications.

As is known today, the activity of Th1 or Th2 cells plays an important role in immune response: Th1 cells promote mainly the cell-mediated immunity, phagocyte-mediated host defense, but also the production of antigen specific IgG2a in mice. In contrast, Th2 cells tend preferentially to support phagocyte independent host-response, IgG1, IgE and IgA immunoglobulin generation.

The Th1 or Th2 basis of an immune response, that is, the differentiation into Th cell subtypes, not only depends on cytokines and the activity of other regulatory molecules (Luger, T. A., Schwarz, T. The role of cytokines and neuro-endocrine hormones in cutaneous immunity and inflammation. *Allergy* (1995) 50: 292-302; Lohoff, M., Gessner, M., Bogdan, C., Roellinghoff, M. The Th1/Th2 paradigm and experimental murine *Leishmaniasis*. *Int. Arch Allergy Immunol.* (1998) 115: 191-202.); the nature of antigen presenting cells and antigen amount used also play an important role. Cytokines are produced transiently by almost all eukaryotic cells and act via specific cell-surface receptors. Indeed, every cell in the skin, after appropriate stimulation, can release such (glyco)protein factors or express their receptors. Most cytokines are pluripotent and can induce each other or else influence the expression of relevant receptors. This allows cytokines to act in synergistic, additive or antagonistic fashion,

within the framework of so-called cytokine cascade (Luger & Schwarz, 1995; *op. cit.*).

The role of different cells in immunoactivation after cutaneous antigen application is as yet incompletely understood (Luger & Schwarz, 1995; *op. cit.*; Lohoff et al., 1998, *op. cit.*). Langerhans cells, located in the suprabasilar skin region, are believed to play the main role in immunopresentation. These cells first bind and process the antigens, then migrate from the epidermis into the lymphatic vessels, and further into the proximal, draining lymph node, bearing the digested antigens with them. During the process Langerhans cells undergo phenotypical and functional alterations and differentiate into (lymphoid) dendritic cells which finally offer the antigens to naive CD4⁺ T cells that have entered the lymph nodes through the high endothelial venules. In contrast, the other two major types of antigen presenting cells in the skin, macrophages and B lymphocytes, first require activation in order to present antigens and stimulate T cells. Antibodies may be presented to T cells by the venular endothelial cells, and perhaps by certain basic cells of the skin as well.

It is clear, for example, that keratinocytes can augment the local inflammation by producing a plethora of proinflammatory cytokines, including IL-1 α , GM-CSF and TNF α (Pastore, S., Fanales-Belaso, E., Abbanesi, C., Chinni, L.M., Giannetti, A., Girolomoni, G. Granulocyte macrophage colony stimulating factor is overproduced by keratinocytes in atopic dermatitis: Implications for sustained dendritic cell activation in the skin. *J. Clin. Invest.* (1997) 99: 3009-3017). Keratinocyte derived cytokines are also critical for the maturation of Langerhans cells into potent antigen presenting cells (Nasir & Gaspari, 1996, *op. cit.*). The extent to which the former cells directly participate in antigen presentation (Kondo & Sauder, 1995, *op. cit.*) is unknown but the production of inhibitory cytokines, such as IL-10, non-functional IL-12 and TGF β , by keratinocytes is an established fact (Nasir & Gaspari, 1996, *op. cit.*).

The fibroblast pool in the skin also contains cellular subsets that are involved in antigen processing. For example, one subset of fibroblasts is recruited selectively by cytokines at the inflammation site in scleroderma (Fries, K.M., Blieden, T., Looney, R. J., Sempowski, G.D., Silvera, M.R., Willis, R. A., Phipps, R. P. Evidence of fibroblast heterogeneity and the role of fibroblast subpopulations in fibrosis. *Clin. Immunol. Immunopathol.* (1994) 72: 283-292.).

It has been reported previously that epicutaneous antigen application produces a different immune response than the more conventional routes of administration through the oral cavity or the nose. For example, after repeated epicutaneous ovalbumin exposure on the skin anti-ovalbumin IgE-s are prominent (Wang et al., 1996, *op. cit.*). Using bovine serum albumin as a model antigen on the skin, an unusually strong IgA production was previously observed (Paul et al., 1997, *op. cit.*), but no consistent picture of the interdependency between the details of epicutaneous antigen presentation and the resulting immune response emerged to date.

Numerous and different cells participate in mounting an immune response against the cutaneously delivered macromolecules. As has been stated above, the approaches taken so far have not led to the establishment of a convincing strategy for generating a protective immune response. This may be due to the fact that the prior art strategies, such as antigen injection, have not assisted in dissecting the immune response obtainable by applying antigens to the skin to an extent that allows for devising a directed and protective immune response. For example, it is known that antigen injection, as any lesion or other kind of skin perturbation, including the presence of chemical irritants, releases various cytokines from the skin (which not only is the heaviest organ in the body but also makes out the major part of the body immune system). This maximizes the strength, but prevents the fine tuning, of cutaneous immune response, which is also sensitive to the nature of antigens used. High impact vaccine delivery profits from this effect.

Material transport across the skin by means of ultradeformable carriers is just the opposite of said high-impact delivery approach, as it reportedly does not affect the skin. It is believed that this is due to the fact that such hydrosensitive, ultradeformable bodies - so called TransfersomesTM (Cevc, 1997, *op. cit.*), penetrate the stratum corneum through 'virtual channels' between corneocytes, adjusted to the shape of the cells (Schätzlein, A., Cevc, G. Non-uniform cellular packing of the stratum corneum and permeability barrier function of intact skin: a high-resolution confocal laser scanning microscopy study using highly deformable vesicles (Transfersomes). *Br. J. Dermatol.* (1998) 138: 583-592.). It was proposed that Transfersomes push the cells in the skin and intercellular lipids apart during the process, preferentially at the sites of

weakest contact. The passages thus generated seem to be approximately 20-30 nm wide, on the average. They cover several percent (~4%) of the skin surface (Schätzlein & Cevc, 1998, *op. cit.*), the draining of adjacent surface not included. This is much more than the normal shunt area (~0.1%), which explains the quantitative differences between the anti-BSA titres measured after antigen administration with ultradeformable carriers (Paul & Cevc, 1995, *op. cit.*) or by using Cholera toxin as an adjuvant (Glenn et al., 1998a, b, *op. cit.*).

Virtual channels in the skin opened by the carriers appear to be sufficiently wide to let the carriers as well as material associated with them pass through the barrier without significantly perturbing the organ. However, repeated insulin delivery across the skin by means of ultradeformable carriers was found not to induce antibodies against the protein (Cevc, G., Gebauer, D., Schätzlein, A. Blume, G. Ultraflexible Vesicles, Transfersomes, Have an Extremely Low Permeation Resistance and Transport Therapeutic Amounts of Insulin Across the Intact Mammalian Skin. *Biochim. Biophys. Acta* (1998) 1368: 201-215.)

The technical problem underlying the present invention was therefore to establish a means that allows for the successful induction of a medically useful transdermal immune response. The solution to said technical problem is achieved by providing the embodiments characterized in the claims.

Accordingly, the present invention relates to a transdermal vaccine comprising (a) a transdermal carrier which is a penetrant, suspended or dispersed in an aqueous solvent, in the form of a minute fluid droplet surrounded by a membrane-like coating of one or several layers of at least two different substances or two different forms of a substance with the tendency to aggregate, said substances or forms of a substance differing by at least the factor of 10 in solubility in a, preferably aqueous, liquid medium, such that the average diameter of homo-aggregates of the more soluble substance or form of the substance or the average diameter of the hetero-aggregates consisting of both said substances or forms of said substance is smaller than the average diameter of homo-aggregates of the less soluble substance, and/or wherein the more soluble component tends to solubilise the penetrating droplet and wherein the content of such component amounts to up to 99 mol-% of the concentration

As regards the above recited values of up to 99%, it is to be noted that values below 50% of the former relative concentration are often used. Even more advantageously values below 40 rel-% or even around and below 30 rel-% are chosen, whereas with the droplets that cannot be solubilised by the more soluble component relative concentrations that exceed the above mentioned ones by the factor of up to 2 are preferred.

In the context of this invention, the term „pathogen“ refers to an entity which through its presence in or on the body leads to or promotes a pathological state which, in principle, is amenable to or could profit from a preventive, curative or adjuvant immunotherapy. This includes pathogens causing microbial diseases such as extracellular bacteria, including pus-forming cocci, such as *Staphylococcus* and *Streptococcus*, gram-negative bacteria, such as *Meningococcus* and *Gonococcus* species, species of *Neisseria*, gram negative bacteria, including enteric organisms such as *E. coli*, *Salmonella*, *Shigella*, *Pseudomonas*, *Diphtheria*, *Bordetella Pertussis*, and gram-positive bacteria (e.g. *Bacillus pestis*, *BCG*), particularly anaerobes, such as the *Clostridium* species (e.g. *Clostridium tetani*, *Clostridium perfringens*, *Clostridium novyi*, *Clostridium septicum*); a number of bacteria and all viruses, which survive and replicate within host cells; this latter group encompasses mycobacteria (e.g. *M. tuberculosis*) and *Listeria monocytogenes*, retro- and adenoviruses, including but not limited to hepatitis virus, (human) immunodeficiency virus, herpes viruses, small-pox, (chicken-pox), influenza, measles, mumps and polio viruses, cytomegalovirus, rhinovirus, etc., and various fungi prospering inside host cells; parasites including animal parasites, such as protozoa and helminths, and ectoparasites, such as ticks

The term “specifically” in combination with “releases” or “induces” denotes the fact that the compound interacts with cells capable of releasing cytokines by a receptor-mediated triggering of this cytokine release or induction. This specific release or induction is in contrast to an unspecific release or induction that is, for example, obtained by an intradermal injection.

The term „(therapeutic) vaccination“ in the context of this invention describes any kind of therapeutic immunization, whether done after the disease has been already established, to improve a clinical situation, or else for the purpose of preventing a

The term „cytokine“, as used in the present invention, denotes cytokines, such as IL-1, IL-2, IL-3, IL4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-11, IL-12, IL-13, IL-14, IL-15, IL-16, IL-17, IL-18, with all subtypes, such as IL-1 α and IL-1 β , tumor necrosis factor (TNF), transforming growth factor (TGF- β and - α), Type I and II interferons (IFN- α 1, IFN- α 2, (IFN- ω), IFN- β , IFN- γ), migration inhibitory factor, MIF, c-kit ligand, granulocyte

macrophage colony stimulating factor (GM-CSF), monocyte macrophage colony stimulating factor (M-CSF), granulocyte colony stimulating factor (G-CSF), chemokines, etc., as well as all functional derivatives of any of these molecules.

Cytokines that mediate natural immunity particularly well include type I interferons (IFN- α and IFN- β), tumor necrosis factor (TNF), interleukin-1 (IL-1 α and IL-1 β), interleukin-6 (IL-6) and leukocytes attracting and activating chemokines. The process relies on antiproliferative (e.g. with IFN-s), pro-inflammatory (e.g. with TNF, IL-1) or co-stimulatory (e.g. with IL-6) action, amongst other. Cytokines which best mediate lymphocyte activation, growth and differentiation include interleukin 2 (IL-2), interleukin-4 (IL-4) and transforming growth factor (TGF). Such cytokines, consequently, not only can affect target growth but, moreover, influence the activation of, and thus the production of other cytokines by, the cells which finally may play a role in therapeutic action.

Cytokines that mediate immune-mediated inflammation, which heavily relies on the cell-mediated response, are interferon-gamma (IFN- γ), lymphotoxin (TNF- β , interleukin-10 (IL-10), interleukin-5 (IL-5), interleukin-12 (IL-12) and, probably, migration inhibition factor. Leukocyte growth and differentiation are most affected by interleukin-3 (IL-3), c-kit ligand, granulocyte-macrophage colony stimulating factor (GM-CSF), macrophage or granulocyte colony stimulating factor (M-CSF or G-CSF) and interleukin-7 (IL-7).

The term „immunoadjuvant“ is used here to describe any substance which supports, augments, stimulates, activates, potentiates or modulates the desired immune response of either cellular or humoral type, specifically in the case of prophylactic treatment by increasing the antigen specific immune response of any kind and in the case of therapeutic treatment often by supporting cell-mediated immunity. This can be achieved by the addition of suitable cytokines, their blends or antagonists, or less directly by the chemical irritation of the skin, when this contributes directly or indirectly to the release of cytokines from the skin or other involved peripheral tissues, or else by catalyzing or promoting the biosynthesis of the molecules in the tissue which then lead to such action, provided that the final outcome is an increased success of vaccination, that is of prophylactic and/or therapeutic action of used antigen. The class of

immunoadjuvants which indirectly contribute to the useful cytokine pool includes small chemical entities with an allergenic potential, such as certain allergenic (metal) ions, including but not limited to LiCl, HgCl₂, molybdenum, acids, bases and other irritating compounds, such as dicyclohexylmethane-4,4'-diisocyanate, dithiocarbamate (diethyldithiocarbamate), 2,4-dinitrochlorobenzene, isoprinosine, isophorone-diisocyanate, levamisole, (phenyl)oxazolone and alike, Swansonine, sizofran, phthalic anhydride, thymopentin, (fatty) alcohols, (fatty) amines, (fatty) ethers, ricin, or other suitable amphiphiles, many surfactants and chemical skin permeation enhancers, as well as derivatives or combinations thereof; furthermore, (low molecular weight) fragments of or derivatives from microbes, including lipopolysaccharides (such as LPS), cord-factor (trehalose-dimycolate) and other polysaccharides attached to membranes, when used in sufficient quantity, acetylmuramyl-alanyl-isoglutamin, and larger fragments of microbes, including bacterial exo- and endotoxins, or enterotoxins, such as cholera toxin and the heat labile toxin (HLT) of *E. coli*, and their macromolecular fragments, such as A-chain derivatives most, if not all, of which seem to possess ADP-ribosylating activity, the high potency immunoadjuvant LT holotoxin, etc., cell-wall skeleton, attenuated bacteria, such as BCG, etc. Less established examples include clostridial toxin, purified protein derivative of *M. tuberculosis*, LT-R192G, Fibronectin-binding protein I of *Streptococcus pyogenes*, outer membrane protein of group B *Neisseria meningitidis* (GBOMP), various other peptidoglycans, etc. Immunoadjuvants, in other words, include molecules that alter the uptake or presentation of antigens, activate or increase the proliferation of antigen specific lymphocytes, or interfere with the dominant control mechanism in the immune response, not just in the skin but also in the other immunocompetent tissues. (The mucosal adjuvant activity of ADP-ribosylating bacterial enterotoxins is a well established and known example for this.) On the other hand, molecules which change the (relative) concentrations of cytokines or other immunoadjuvants, such as anti-immunoadjuvant antibodies or other agonists or antagonists of immunoadjuvants, also are immunoadjuvants in the sense of this invention. The same is true for molecules which affect lymphocyte homing, such as various selectins (LECAMS, e.g., various CD62-s), GlyCAM-1, MadCAM-1, VCAM-1, ICAM-1, hyaluronate, etc., and other chemokines, such as RANTES or MCP-1. Endogenous group of immunoadjuvant furthermore comprises histamines, transfer factor, tuftsin, etc. As many of the above mentioned immunoadjuvants do not have sufficient potency to ensure the desired

effect after the non-invasive immunization at too low, and sometimes too high, concentration or on their own, the functional definition of an adjuvant used in this work includes a fortiori sufficient and such modulation of cytokine concentration and distribution pattern in the body that results in mounting the desired therapeutic or prophylactic immune response. If required to gain clarity said modulation and its extent must be determined in a dedicated experiment, in which the specific cytokine levels are determined, for example.

„Immunoadjuvant manipulation“ denotes a non-chemical treatment of the skin, such as skin rubbing, pressing, heating, exposing to an electrical or mechanical, e.g., ultrasound, field, etc., or even an injection of a non-immunogenic formulation in the skin, provided that such treatment releases immunoadjuvant compounds from the skin or other peripheral immuno-active tissues or else reduces the concentration/duration of action of antagonists to the desired vaccination. However, the term „immunoadjuvant manipulation“ also denotes, in accordance with the present invention, a pre-treatment of the skin with immunomodulators and/or cytokines and/or cytokine releasing factors, like, inter alia, histamine.

The term „immunogen“ denotes a hapten coupled to an immunological carrier or an antigen, free or associated with a carrier, which is capable of inducing an immune response.

„Immuno-tolerance“ denotes the lack or, more generally, the reduction of an undesired immune response to an antigen.

Th1 (T-helper cell type I) related antibodies include IgG2a, IgG2b and IgG3.

Th2 (T-helper cell type II) related antibodies comprise the classes of IgG1, IgG4 and IgE.

The term “two forms of a substance” in connection with this invention means two ionization states or salt forms of the same substance, two different complexes of such substance, etc.

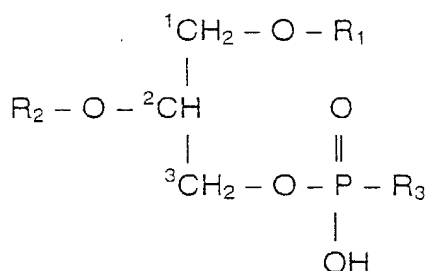
„Penetration“ describes a non-diffusive motion of relatively large entities across a barrier. This process typically relies on the penetrant adaptation to the otherwise confining pores in the barrier and also may involve a penetration induced decrease in the barrier resistance, such as pore widening or channel opening; the process does not depend, however, primarily on the penetrant concentration gradient across the barrier.

A penetrant, consequently, is an entity comprising a single molecule or an arrangement of molecules too big to permeate through a barrier but capable to cross the barrier owing to the penetrants adaptability to the shape and/or diameter of the otherwise confining passages (pores) of a barrier. This adaptability is seen from the fact, for example, that penetrants more than twice bigger than the pore diameter will cross the bilayer without being fragmented down to the pore size. A permeant, on the other hand, is an entity that can permeate through the semi-permeable barrier, such as the skin. A penetrant in an external field experiences a driving force proportional to the nominal penetrant size and to the applied field, which may occur naturally. Such a force, which on the intact, non-occluded skin is believed to originate from the water concentration gradient across the stratum corneum, can result in a penetrant motion through the barrier, including the skin, if the force is strong enough either to deform the penetrant or else to widen the passages in the barrier sufficiently to elude the problem of size exclusion, or both.

For further definitions, especially such pertaining to the penetrants in terms of complex body deformability, the corresponding mechanism of action, lists of interesting penetrant ingredients or selected agents it is referred to the issued or pending patents (DE 41 07 152, PCT/EP91/01596, PCT/EP96/04526, DE 44 47 287). Detailed information relevant for the manufacturing process and penetrant loading with the

antigenic (macro)molecules and/or immunoadjuvants, which are too big to permeate through the barrier, can be found in international patent application PCT/EP98/06750.

Typically, the less soluble amongst the aggregating substances forming a carrier is a lipid or lipid-like material, especially a polar lipid, whereas the substance which is more soluble in the suspending liquid and which increases the droplet adaptability belongs to surfactants or else has surfactant-like properties. The former ingredient, typically, is a lipid or lipid-like material from a biological source or a corresponding synthetic lipid or any of its modifications, such lipid often belonging to the class of pure phospholipids with the chemical formula



where R_1 and R_2 is an aliphatic chain, typically a C_{10-20} -acyl, or -alkyl or partly unsaturated fatty acid residue, in particular, an oleoyl-, palmitoeloyl-, elaidoyl-, linoleyl-, linolenyl-, linolenoyl-, arachidoyl-, vaccinyl-, lauroyl-, myristoyl-, palmitoyl-, and stearoyl chain, and where R_3 is hydrogen, 2-trimethylamino-1-ethyl, 2-amino-1-ethyl, C_{1-4} -alkyl, C_{1-5} -alkyl substituted with carboxy, C_{2-5} -alkyl substituted with hydroxy, C_{2-5} -alkyl substituted with carboxy and hydroxy, or C_{2-5} -alkyl substituted with carboxy and amino, inositol, sphingosine, or salts of said substances, said lipid comprising also glycerides, isoprenoid lipids, steroids, sterines or sterols, of sulphur- or carbohydrate-containing lipids, or any other bilayer forming lipids, in particular half-protonated fluid fatty acids, and preferably is selected from the group of phosphatidylcholines, phosphatidylethanolamines, phosphatidylglycerols, phosphatidylinositols, phosphatidic acids, phosphatidylserines, sphingomyelins or other sphingophospholipids, glycosphingolipids (including cerebrosides, ceramidepolyhexosides, sulphatides, sphingoplasmalogens), gangliosides or other glycolipids or synthetic lipids, in particular with or else with corresponding sphingosine derivatives, or any other glycolipids, whereby two similar or different chains can be

physicochemical states of the same (polar) lipid mixed together in a membrane will produce a highly deformable carrier satisfying the conditions of this work.

More general information on lipid suspensions can be found in handbook dealing with 'Liposomes' (Gregoriadis, G., Hrsg., CRC Press, Boca Raton, FL, Vols 1-3, 1987), in the book 'Liposomes as drug carriers' (Gregoriadis, G., Hrsg., John Wiley & Sons, New York, 1988), or in the laboratory manual 'Liposomes. A Practical Approach' (New, R., Oxford-Press, 1989). The properties of phospholipids which can be used conveniently to prepare bio-compatible immunopenetrants are reviewed in 'Phospholipids Handbook' (Cevc, G., ed., Dekker, New York, 1995).

It may be convenient to adjust the pH value of a formulation immediately after preparation or just before its application. Such an adjustment should prevent the deterioration of individual system components and/or drug carriers under the conditions of initial pH without sacrificing physiological compatibility. To neutralize a penetrant suspensions it is reasonable to use biocompatible acids or bases to prepare buffers with a pH value between 3 and 12, frequently between 5 and 9 and most often between 6 and 8. Physiologically acceptable acids are, for example, diluted aqueous solutions of mineral acids, such as hydrochloric acid, sulphuric acid, or phosphoric acid, or organic acids, such as carboxyalkane acids, e.g. acetic acid. Physiologically acceptable bases are, for example, diluted sodium hydroxide, suitably ionized phosphoric acid, etc.

If required, immunogen suspension can be diluted or concentrated (e.g. by ultracentrifugation or ultra-filtration) before the application; additives can also be given into the suspension at this time or before. The additives are often selected amongst substances that reduce the formulation sensitivity to ambient stress, including microbicides, anti-oxidants, antagonists of undesired enzyme action, in case cryo-preservants, thickening agents, etc. However, after any system manipulation, the carrier characteristics should be checked and, if required, readjusted.

In accordance with the present invention it was surprisingly found that macromolecular antigens associated with ultradeformable lipid aggregates described herein (immunopenetrants) can cross artificial porous barriers as well as the skin, despite the fact that

immune response, or else will result in the gradual development of tolerance against the antigens or in the specific promotion of cell-mediated immunity.

In accordance with the present invention, it was also found that the outcome of noninvasive transcutaneous vaccination is strongly affected by the immuno-penetrant (antigen carrier) composition. Using antigens of different purity, unexpectedly, resulted in vastly different immune response. This was reflected in the observation that organisms with a similar overall titre revealed diverse levels of protection, probably due to the different final antigen isotype patterns.

Furthermore, the addition of a conventional, low molecular weight immunoadjuvant, monophosphoryl lipid A, not only made the result of epicutaneous immunization more robust, as documented by the smaller standard deviation in the measured antibody titres published before. Using this immunoadjuvant in immuno-carriers also, unexpectedly and contrary to previous experience obtained in mice, increased the secretion of IgG2b, and less strongly of IgG2a, but did not enhance IgA production. As the presence of IgG1, which is a Th1-like immunoglobulin, is inferred to be essential for, at least murine, protection against the tetanus toxin, the role of lipid A or bacterial antigens was thus revealed for the first time. For the future medical and commercial use of teachings disclosed in this invention it is important to realize that a high (specific) antibody titre does not necessarily imply a good protection result; to achieve the desired and sufficient protection the right kind and relative amount of certain antibody isotypes is required, such that will give prevalently Th1- or Th2-type of immune response (see previous discussion), as the case should be.

Basic formulations suitable for achieving the desired goals are known in the art: see, e.g., DE 41 07 152, PCT/EP91/01596, PCT/EP96/04526, DE 44 47 287, for more detailed or complementary information. The vaccine of this invention is not useful just for prophylactic or therapeutic vaccination but, moreover, is applicable for the treatment of allergy and for obtaining immunity against microbes, including extracellular and intercellular bacteria, viruses and parasites in the human and veterinary medicine.

In combination with the above mentioned penetrants, an antigen, such as an immunoactive substance, is transported across the barrier in form of a physical or a chemical complex with the former.

In order to profit from the pool of cytokines residing in the skin, a particularly useful method of vaccination is proposed in which an immunogen is applied on the skin after pre-treating the organ by an immunoadjuvant manipulation as defined before.

It is particularly advantageous to use the readings from the above mentioned local immune response to a patch assessment for optimizing the details and the course of further allergen administration, and thus to positively affect the outcome of therapeutic or prophylactic vaccination. It is believed that such an approach could be used advantageously to reach or improve immuno-tolerance of the tested subject to an applied allergen.

If primary immunization is done invasively, typically by using a subcutaneous injection or some other suitable skin barrier perforating/destructing method, one expects to obtain high IgM levels but the subsequent, booster immunizations may then be done non-invasively as described in this invention.

Finally, several optimization methods are proposed which can be used to improve immunogens and vaccination based on highly deformable penetrants. Preferred is a method wherein the flux of penetrants associated with an immunogen through the various pores in a well-defined barrier is determined as a function of suitable driving force or pressure acting across the barrier and the data are then conveniently described by a characteristic curve which, in turn, is employed to optimize the formulation or application further. Its core is the determination of the flux of immuno-penetrants through the pores in a well-defined barrier as a function of suitable driving force or pressure, which acts across the barrier, and the resulting data analysis in terms of a characteristic curve which, in turn, can be employed to optimize the formulation or application further, based on comparison of different data sets. This includes comparison with the results pertaining to the immunogen-free penetrant suspensions of known skin penetration capability, reported for example by Cevc et al., (1998, *op. cit.*). In a complementary, preferred, embodiment various combinations of

The invention in one further preferred embodiment relates to a vaccine wherein total weight of droplets in the formulation for the use on human or animal skin is 0.01 weight-% (w-%) to 40 w-% of total mass, in particular between 0.1 w-% and 30 w-%, and most preferably between 5 w-% and 20 w-%.

In another preferred embodiment of the vaccine according to the present invention the formulation further comprises (da) a low molecular weight chemical irritant, and/or (db) a low molecular weight compound from a pathogen or a fragment or a derivative thereof.

In one more preferred embodiment of the vaccine according to the present invention the compound displaying anti-cytokine activity is an anti-cytokine antibody or the corresponding active fragment, a derivative or an analogue thereof.

The term "active fragment or derivative thereof" in this connection means that the above-recited activity is essentially maintained or mimicked by the substance used.

In another particularly preferred embodiment of the vaccine according to the present invention said pathogen is selected from extracellular bacteria, including pus-forming cocci, such as *Staphylococcus* and *Streptococcus*, gram-negative bacteria, such as *Meningococcus* and *Gonococcus* species, species of *Neisseria*, gram negative bacteria, including enteric organisms such as *E. coli*, *Salmonella*, *Shigella*, *Pseudomonas*, *Diphtheria*, *Bordetella Pertussis*, and gram-positive bacteria (e.g. *Bacillus pestis*, *BCG*), particularly anaerobes, such as the *Clostridium* species (e.g. *Clostridium tetani*, *Clostridium perfringens*, *Clostridium novyi*, *Clostridium septicum*), bacteria and viruses, which survive and replicate within host cells, comprising mycobacteria (e.g. *M. tuberculosis*) and *Listeria monocytogenes*, retro- and adenoviruses, including hepatitis virus, (human) immunodeficiency virus, herpes viruses, small-pox (chicken-pox), influenza, measles, mumps and polio viruses, cytomegalovirus, rhinovirus, etc., and fungi prospering inside host cells, parasites including animal parasites, such as protozoa and helminths, and ectoparasites, such as ticks and mites, or *Brucella* species (e.g. *B. melitensis*, *B. abortus*, *B. suis*, *B. canis*, *B. neotomae*, *B. ovis*), the causative agent for cholera (e.g. *Vibrio cholerae*), *Haemophilus* species like *H. actinomycescomitans*, *H. pleuropneumoniae*, as well as pathogens triggering paratyphoid, plague, rabies, tetanus and rubella diseases and pathogens that cause various neoplasiae, auto-immune diseases or are related to other pathological states of the animal or human body which do not necessarily result from pathogen infections.

In a preferred embodiment of the vaccine according to the present invention the allergen is of xenogenic or endogenic origin, derived from a microorganism, an animal or a plant which leads to an acute hypersensitivity reaction of the body exposed to the allergen, many such allergens stemming from mite, pollen, animal hair or skin debris, or belonging to the group of man made and/or irritating inorganic substances, or to such parts or components of the human body which were incorrectly processed by or exposed to the body immune system.

In a further preferred embodiment of the vaccine according to the present invention the concentration of each compound displaying cytokine activity used is selected to be up to 1000 times higher than the concentration optimum established in the corresponding tests with the antigen and immunoadjuvant chosen, performed by subcutaneously injecting the formulation or performing the tests *in vitro*, and preferably is up to 100x, more often up to 50x and even better up to 20x higher.

In a different preferred embodiment of the vaccine according to the present invention the pathogen extract or compound is a lipopolysaccharide, cord-factor (trehalose-dimycolate), muramyl dipeptide, or another (poly)saccharide or (poly)peptide identical to or resembling an immunologically active part of a membrane of a pathogen; an extract of a pathogen, including bacterial exo- and endotoxins, preferably cholera toxin or the heat labile toxin (HLT) of *E. coli*, an A-chain derivative, a component with an ADP-ribosylating activity, a peptidoglycane, a clostridial toxin, or a purified protein derivative of *M. tuberculosis*, LT-R192G, Fibronectin-binding protein I of *Streptococcus pyogenes*, or outer membrane protein of group B *Neisseria meningitidis* (GBOMP); or bacterial or viral nucleic acids such as oligonucleotides containing unmethylated CpG dinucleotides.

In a particularly preferred embodiment of the present invention said lipopolysaccharide is lipid A or a derivative and modification thereof, such as monophosphoryl lipid A, or its analogue, such as a fatty derivative of saccharose.

In another particularly preferred embodiment of the vaccine according to the present invention the concentration of the pathogen compound derived from a pathogen is between 10x lower and up to 1000x higher than that otherwise used with the corresponding injected formulations employing similar antigen, the epicutaneously administered immunoadjuvant concentration more often differing from the injected immunoadjuvant concentration by the factor between 0.5 and 100, or better, by the factor between 1 and 50, and best between 2 and 25.

In still another particularly preferred embodiment of the vaccine according to the present invention the low molecular weight irritant is selected from the classes of allergenic metal ions, acids, bases, irritating fluids, (fatty-) alcohols, (fatty-) amines,

(fatty-) ethers, (fatty-) sulphonates, -phosphates, etc., or other suitable solvents or amphiphiles, or from the group of surfactant-like molecules, often with the skin permeation enhancing capability, as well as derivatives or combinations thereof.

In a preferred embodiment of the vaccine according to present invention the concentration of a low molecular weight irritant is chosen to be by at least the factor of 2, more often by the factor of 5, and even better by the factor of 10 or more, below the concentration which in independent tests on the same or comparable subject is deemed to be unacceptable owing to the local irritation, as assessed by the methods and standards commonly used to test such an irritant.

In a further particularly preferred embodiment of the vaccine according to the present invention the allergen belongs to the class of inhalation allergens, including various pollen, spores, bits of animal hair, skin, feather, natural and synthetic textiles, wheat, (house) dust, including mite; food and drug allergens; contact allergens; injection, invasion and depot allergens, such as various (gastrointestine-resident) worms, echinococci, trichines, etc., parts of implantation material, etc..

In a preferred embodiment of the vaccine according to the present invention the applied dose of an antigen differs by the factor of 0.1 to 100 from the dose which otherwise would have to be injected in the process of immunization, but more often is in the range between 0.5 to 50, even better between 1 and 20 and ideally is less than 10x higher than that used with an injection.

In another preferred embodiment of the vaccine according to the present invention the applied penetrant dose is between 0.1 mg cm^{-2} and 15 mg cm^{-2} , even more often is in the range 0.5 mg cm^{-2} and 10 mg cm^{-2} , and preferably is between 1 mg cm^{-2} and 5 mg cm^{-2} . It may also be advantageous to use different administration areas to control the applied immunogen dose, using easily accessible or sheltered body areas (such as the chest or back regions, arms, lateral side of the neck, e.g. behind the ears, or even in the scalp region) for the purpose.

In a different preferred embodiment of the vaccine according to the present invention said antigen is a pure or purified antigen. The use of highly purified antigens in the

vaccine of the invention has turned out to be particularly advantageous for the generation of a protective immuno response.

The present invention further relates to a kit comprising, in a bottled or otherwise packaged form, at least one dose of the vaccine.

In a preferred embodiment according to the present invention the kit comprises at least one injectable dose of the antigen described above.

The present invention further relates to a method for generating a protective immune response on a mammal comprising vaccinating said mammal with a vaccine as described above.

In another preferred embodiment of the method according to the present invention different treatment areas are selected to control the applied immunogen dose and the outcome of therapeutic vaccination.

In one more preferred embodiment of the method according to the present invention a suspension of antigen-free penetrants is loaded with the antigen to be associated therewith during the day prior to an administration, preferably 360 min, more preferably 60 min and even more preferably 30 min before the administration of resulting formulation on the skin.

In a different preferred embodiment of the method according to the present invention the vaccine of the present invention is applied on the skin after pre-treating the organ by an immunoadjuvant manipulation, said manipulation comprising, for example, skin rubbing, pressing, heating, exposing to an electrical or mechanical, e.g. ultrasound field, etc., or injecting a non-immunogenic formulation in the skin, provided that any such treatment releases immunoadjuvant compounds from the skin or other peripheral immuno-active tissues or else reduces the concentration/duration of action of antagonists to the desired vaccination.

In a preferred embodiment of the method according to the present invention immunogen is applied in a non-occlusive patch. This embodiment can also be used for

the purpose of assessing the skin reaction to an epicutaneously administered immunogen in the penetrant suspension, to which the former, at least originally, is allergic and which thus gives rise to an acute local hypersensitivity reaction, as seen, for example from the resulting flare, irritation, etc.

In another preferred embodiment of the method according to the present invention at least one dose of vaccine is administered.

This embodiment of the method of the invention includes the repeated administration of the vaccine of the invention. Repeated administration includes repeated administration on the skin or one or more administrations on the skin in combination with, e.g., parenteral administrations. In this connection, the kit of the invention may be advantageously used that comprises one or more containers or ampoules comprising the vaccine of the invention.

In a particularly preferred embodiment of the method according to the present invention said vaccine is administered as a booster vaccination.

In a most preferred embodiment of the method according to the present invention the primary immunization is done invasively, typically using a subcutaneous injection or some other suitable skin barrier perforating/destructing method, and the at least one subsequent, booster immunization is done non-invasively.

In a preferred embodiment of the method according to the present invention the vaccine is applied between 2 and 10, preferably between 2 and 7, even more preferably up to 5 and most preferably up to 3 times, when a non-allergenic antigen is used, or such a number of times, in the case of allergens, as is required either to achieve the desired immuno-tolerance, determined according to a suitable assessment method, or else to deem the effort as having failed.

In a particularly preferred embodiment of the method according to the present invention the time interval between the subsequent vaccinations is chosen to be between 2 weeks and 5 years, often between 1 month and up to 3 years, more frequently between 2 months and 1.5 years. In a further preferred embodiment,

repeated immunogen administration is advocated to maximize the final effect of a therapeutic vaccination. It is proposed to use between 2 and 10, often between 2 and 7, more typically up to 5 and most preferred up to 3 immunizations, when a non-allergenic antigen is used, or such a number of times, in the case of allergens, as is required either to achieve the desired immuno-tolerance, determined as described above or another suitable assessment method, or else to deem the effort as having failed. The time interval between subsequent vaccinations should preferably be between 2 weeks and 5 years, often between 1 month and up to 3 years, more frequently between 2 months and 1.5 years, when a subject is being immunized for the first time. Rodents, such as mice and rabbits are advantageously immunized in 2 weeks interval, primates, e.g., monkeys and often humans, need a booster vaccination in 3-6 months interval.

In a preferred embodiment of the method according to the present invention the flux of penetrants that carry an immunogen through the various pores in a well-defined barrier is determined as a function of a suitable driving force or a pressure acting across the barrier and the data are then conveniently described by a characteristic curve which, in turn, is employed to optimize the formulation or application further.

The invention finally relates to the use of the transdermal carrier, the compound which specifically releases or specifically induces cytokine or anti-cytokine activity or exerts such an activity, the antigen or allergen, and optionally an extract or a compound from a microorganism or a fragment or a derivative thereof, and/or a low molecular weight chemical irritant as defined hereinbefore for the preparation of a vaccine for inducing a protective or tolerogenic immune response.

~~The figures show:~~

~~Figure 1 gives the data on survival of animals immunized epicutaneously with mixed micelles or Transfersomes loaded with TT, to illustrate aggregate size (stability) effect, since the over-destabilized Transfersomes normally disintegrate into the mixed lipid micelles.~~

The figures show:

Figure 1: Mixed micelles versus Transfersomes. The figure gives the data on survival of animals immunised epicutaneously with mixed micelles or Transfersomes loaded with purified TT, to illustrate aggregate size (stability) effect, since the over-destabilised Transfersomes normally disintegrate into the mixed lipid micelles.

Figure 2: Liposomes versus Transfersomes. A comparison is made between the immune response to conventional lipid vesicles (liposomes) and ultradeformable lipid vesicles (Transfersomes) carrying purified TT and applied on the skin. The information on corresponding specific antibody concentrations in serum (expressed as absorbance) is given in the upper panel.

Figure 3: Antigen dose effect. The figure illustrates the effect of increasing antigen dose on the outcome of epicutaneous immunisation by means of Transfersomes from SPC:NaChol (3.75:1) loaded with antigen and monophosphoryl lipid A (LA). The results are expressed as absorbance change, antibody titre, or animal survival, together with the corresponding specific antibody isotyping data. Antigen doses were 10, 20, 40 and 80 μ g. 6 animals per each group except for No Ag (4 animals) were used.

Figure 4: Antigen purity effect. The figure highlights the effect of antigen purity on the result of epicutaneous immunisation with 80 μ g tetanus toxoid and monophosphoryl lipid A (LA) in Transfersomes from SPC:NaCh (3.75:1), including information on time dependence of animal survival. All data were obtained after the 2nd boost + 7 days.

Figure 5: Epicutaneous versus subcutaneous immunization. The figure compares the outcome of repeated invasive (subcutaneous) and non-invasive (epicutaneous) immunisation by means of TT in Transfersomes, including animal

survival, serum concentration (in terms of absorbance), specific antibody titre, and antibody distribution pattern values.

Figure 6: Pre-injection effect. The figure illustrates the effect of skin pre-treatment (non-specific challenge) on the immune response following Transfersome (SPC:Tw-80 1:1) mediated TT (40 μ g) delivery across the skin. Mice in the preinjection groups were injected 24 hours before the application of 40 μ g antigen. 0.1 ml each of saline (pre-S), 10% SPC:NaCh 4.5:1 empty Transfersomes (Pre-empty Tfs), and incomplete Freund's adjuvant were used for pre-injection. All mice in this experiment were challenged with 50 times LD50 dose of toxin 7 days after the second boost. It means (ec) epicutaneous, (sc) subcutaneous, and (Tfs) Transfersomes.

Figure 7: Adjuvant effect: for example monophosphoryl lipid A. The figure focuses on adjuvant effect of a relatively low-molecular weight immuno-stimulator, monophosphoryl Lipid A (LA), delivered across intact skin together with TT in Transfersomes.

Figure 8: Adjuvant effect: for example cytokine IL-12. The figure demonstrates the immuno-adjuvancy of a cytokine, interleukin-12 (IL-12) transported across the skin (ec) together with TT by means of Transfersomes from SPC:NaCh.

Figure 9: Immunomodulant effect, for example cytokines. The figure deals with the immuno-modulation by various cytokines of the murine response against impure tetanus toxoid (TT) antigen delivered in Transfersomes non-invasively through the skin. Serum was collected for the assay on the 7th day after 2nd boost. No protection was observed in any of the groups.

Figure 10: Immunoadjuvant effect: for example cholera toxin (CT). The figure presents experimental evidence for the immune response stimulation of mice treated on the skin by pure tetanus toxoid (TT) in Transfersomes (SPC:NaCh 3.75:1), when the carriers also include 10 μ g cholera toxin (CT) to support the

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specific antibody production, and thus animal protection against an otherwise lethal challenge by the tetanus toxin. 4-6 animals per group were used. The asterisc indicates 1 paralyzed mouse out of 4.

Figure 11 Adjuvant effect: for example heat labile toxin (HLT) from E.coli. The figure illustrates the use of heat labile toxin from E. coli as an immuno-
adjuvant.

Figure 12: Histamine effect: on anti-tetanus titer and survival after immunization with Transfersomes on the skin. The figure illustrates the immuno-modulating effect of local skin pre-treatment with histamine in combination with transdermal antigen application with Transfersomes.

Figure 13: Subcutaneous priming: effect on anti-tetanus titer and survival after epicutaneous boosts. The figure demonstrates the effect of subcutaneous priming on anti-tetanus titer and on the survival of epicutaneously vaccinated hosts.

Figure 14: Bi-valent vaccines: Anti-Tetanus and anti-Cholera response to the administration of both antigens together in Transfersomes on the skin. The figure shows the effect of bi-valent vaccination with Tetanus Toxoid nad Cholera Toxin used as antigens.

~~Figure 10 presents experimental evidence for the immune response stimulation of mice treated on the skin by TT in Transfersomes, when the carriers also include cholera toxin (CT) to support the specific antibody production, and thus animal protection against an otherwise lethal challenge by the tetanus toxin.~~

Figure 11 illustrates the use of heat labile toxin from *E. coli* as an immuno-adjuvant.

Figure 12 illustrates the immuno-modulating effect of local skin pre-treatment with histamine in combination with transdermal antigen application with Transfersomes.

Figure 13 demonstrates the effect of subcutaneous priming on anti-tetanus titer and on the survival of epicutaneously vaccinated hosts.

~~Figure 14 show the effect of bi-valent vaccination with Tetanus Toxoid and Cholera Toxin used as antigens.~~

The examples illustrate but do not define the limits of the invention.

Général experimental set-up and sample preparation

Mice of Swiss albino strain (18-20 g) were obtained from The National Institute of Nutrition (Hyderabad, India). They were 8 to 12 weeks old at the time of first immunization and were normally kept in suspension cages in groups of 4 to 6. The animals had free access to standard chow and water. One day prior to an immunization, the application area on murine back was shaved carefully. The antigen was administered with a high precision pipette on the skin surface and left to dry out partially. To prevent immunogen abrasion, the animals were transferred into individual cages in which they were kept for 18 hours following each epicutaneous material administration.

General anesthesia was used to keep the test animals stress free and quiet during manipulations, including immunization. An injection of a mixture of Ketavet and Rompun (0.3 mL per mouse of an isotonic NaCl solution containing 0.0071 % Rompun

(Bayer, Leverkusen, Germany) and 14.3 mg/mL Ketavet (Parke-Davis, Rochester, N.Y) into the peritoneal cavity was used for the purpose. This typically kept the animals asleep for app. 2 hours.

Immunogens

Ultradeformable immuno-carriers, or immuno-penetrants (immuno-Transfersomes), studied in this work, typically had the form of (oligo)bilayer vesicles. They contained biocompatible (phospho)lipids, such as phosphatidylcholine, and (bio)surfactants, such as sodium cholate or polysorbate (Tween 80), different compositions maintaining the high aggregate deformability being possible. Additional ingredients were monophosphoryl lipid A, with a versatile immunoadjuvant activity, and antigens, as required and specified.

Conventional vesicles, liposomes, comprised soy phosphatidylcholine (SPC; Nattermann Phospholipids, Rhone-Poulenc Rorer, Cologne, Germany) and were prepared as described as follows. An organic lipid solution with or without the adjuvant monophosphoryl lipid A (MLA) at 0.04 mol-% relative to SPC was first dried under vacuum (10 Pa, overnight). The resulting lipid film was hydrated with a solution of tetanus toxoid (2.0 mg/mL; Accurate antibodies, NY, USA) in phosphate buffer (pH = 6.5) to get a 10 wt-% lipid suspension. Crude suspension of lipid vesicles was extruded through the series of polycarbonate membranes with 800 nm, 400 nm, and 200 nm pores, to narrow down the final vesicle size distribution.

Highly deformable vesicles, Transfersomes, were prepared as described earlier (Paul et al., 1995 *op. cit.*). In short, an ethanolic SPC solution was mixed with sodium cholate (Merck, Darmstadt, Germany) (3.75/1 mol/mol) and the adjuvant, if required. The mixture was dispersed in 10 mM phosphate buffer (pH = 6.5). This was done with tetanus toxoid present in the solution to give between 0.25 mg and 2.0 mg protein per 1 mL of suspension, as required. Vesicle suspension was then frozen and thawed three times. Subsequently, the formulation was passed through a micro-porous filter (200 nm; Poretics, CA) under pressure. To check the reproducibility of vesicle manufacturing, the optical density at 400 nm was measured with each preparation and confirmed to be approximately constant.

By varying surfactant-to-lipid ratio the vesicular aggregate deformability was controlled, up to the concentration at which membranes became unstable, owing to the high surfactant concentration, and reverted into a micellar form. Lipid vesicles without the surfactant added, which are commonly known as liposomes and have at least 10x less flexible membranes than Transfersomes, were used as negative controls.

Total lipid concentration was typically 10 w-%, unless stated otherwise. Antigen concentration was typically, but not necessarily, of the order of 1 mg/mL. A buffer containing microbicide provided the bulk phase. For other suitable compositions the expert is explicitly referred to other publications and patents from our laboratory.

Immunizations were done with different formulations, including the ultradeformable vesicles without antigens; such vesicles then contained the tetanus toxoid (with or without lipid A) and free immunogen. Each formulation was tested on six mice, unless stated otherwise.

In the case of subcutaneous immunization, 40 μ g of immunogen was injected per mouse. For a non-invasive administration, tetanus toxoid doses between 1 μ g and 80 μ g, associated with different carriers, were administered per mouse on the intact skin of upper dorsum. All non-injected formulations were applied with a high precision pipette and left to dry; during this period mice were kept in separate cages to minimize the applied material abrasion, such as might result from the rubbing of the murine backs on each other. Animals were boosted every two weeks, that is on days 14 and 28; the total immunization scheme thus consisted of three doses, and comprised a prime and two boosts.

Animals were bled retro-orbitally on the days 7, 21 and 35. The collected blood was first allowed to clot. After a brief centrifugation in a micro-centrifuge the serum was separated, de-complemented at 56 °C for 30 min, and then stored at -20 °C, until the total antibody concentration and the specific antibody isotypes was determined.

Absorbency measurements were done using standard UV-vis spectrometer.

Challenge with antigen (the tetanus toxin) in vivo. On the day 35, test animals

Non-protected mice showed signs of paralysis after 24 hours resulting in death, after 36 hours, at latest. Animals which developed no symptoms of paralysis or other anomaly over a 4 days period following the challenge were deemed immune against tetanus.

Examples 1-2:

Highly deformable vesicles (TransfersomesTM: IDEA):

12.6 mg sodium cholate (NaChol)

0.9 mL phosphate buffer, 10 mM, pH 6.5

(Mixed lipid) Micelles:

35 mg sodium cholate (NaChol)

0.9 mL phosphate buffer, 10 mM, pH 6.5

Tetanus toxoid (2 mg/mL; Accurate Antibodies) used at the dose of

40 μg (20 μL) or 80 μg (40 μL) TT per mouse and immunization

Application area: 1 cm² or 2 cm² for 40 µg or 80 µg TT per mouse on the upper dorsum.

To test the effect of formulation stability on the immunological properties of various, epicutaneously administered formulations, two kind of aggregates were prepared: relatively large vesicles (diameter between 100 nm and 200 nm) and relatively small micelles (diameter below 50 nm). The latter were chosen in the expectation that under suboptimal conditions (owing to the lipid degradation or inappropriate aggregate composition) the latter may arise from the former.

Antibody titres, as reflected in the serum absorbency at 492 nm, are shown in figure 1. They show that mixed lipid micelles are less efficient antigen carriers than ultradeformable mixed lipid vesicles (Tfs) loaded with the same amount of TT. Mixed micelles containing less potent detergents (with lesser skin permeation enhancing capability) were even less efficient immune response mediators.

Animal protection data reveal a similar trend, as is seen in lower panel of figure 1.

Examples 3-4:

Aggregate deformability effect

Conventional lipid vesicles (liposomes):

- 100 mg phosphatidylcholine from soy bean (SPC)
- 0.4 mol-% monophosphoryl Lipid A (MLA) relative to SPC
- 0.5 mL phosphate buffer, 10 mM, pH 6.5
- 2 mg/mL tetanus toxoid (Accurate Antibodies)

Highly deformable vesicles (Transfersomes™):

- 87.4 mg phosphatidylcholine from soy bean (SPC)
- 12.6 mg sodium cholate (NaChol)
- 0.04 mol-% monophosphoryl Lipid A (MLA) relative to SPC
- 0.9 mL phosphate buffer, 10 mM, pH 6.5
- 0.1 mL ethanol

Tetanus toxoid used at the dose of 40 µg or 80 µg TT/ mouse/ immunization

Application area: 1 cm² or 2 cm² for 40 µg or 80 µg TT/ mouse/ immunization on the upper dorsum.

Results obtained with the conventional vesicles differ from the data measured with highly deformable vesicles: simple liposomes, which do not cross the narrow pores in a barrier also do not elicit a substantial antibody titre. Conversely, the vesicles with a highly flexible and deformable, and thus better adaptable, membrane which were shown separately to move through the narrow pores in a barrier with greater ease, generate an appreciable quantity of antibody when applied on intact skin, according to the results of serum absorbency measurements (cf. figure 2).

Examples 5-10:

Antigen dose effect

Highly deformable vesicles:

- 86.3 mg phosphatidylcholine from soy bean (SPC)
- 13.7 mg sodium cholate (NaChol)
- 0.04 mol-% monophosphoryl Lipid A (MLA) relative to SPC
- 0.9 mL phosphate buffer, 10 mM, pH 6.5
- 0.1 mL ethanol

Tetanus toxoid (TT: Accurate Antibodies, New York, USA) concentration:

- empty, 0.25 mg/mL, 0.5 mg/mL, 1 mg/mL, 2 mg/mL, giving raise to
- 0 µg, 10 µg, 20 µg, 40 µg or 80 µg TT/ mouse/ immunization

Application area: 1 cm² for 0 µg, 10 µg, 20 µg, 40 µg and 2 cm² for 80 µg TT/mouse/immunization on the upper dorsum.

The results of this experimental series are illustrated in figure 3. It clearly shows the increase in immune response to epicutaneously administered tetanus toxoid in ultradeformable carriers with increasing TT dose. This is reflected in serum absorbency (up to the dose of 20 µg/immunization), in specific antibody titre (up to the dose of 40 µg/immunization), and in the survival data (which do not saturate for doses up to 80 µg/immunization).

Less clarity is found in isotype distribution pattern, except for IgG1 (with a strong indication for the response saturation) and for IgG2b (perhaps, with the saturation between 40 μ g and 80 μ g per immunization). IgM shows dose dependence similar to that of IgG1. The picture obtained for IgG2a is confusing.

Examples 11-13:

Antigen purity effect

Highly deformable vesicles:

as described with examples 5-10 (except in that the group treated with impure TT did not receive immunoadjuvant lipid A)

Tetanus toxoid: 2 mg/mL, corresponding to 80 μ g TT per mouse/ immunization

Application area: 2 cm² on the upper dorsum.

Antigen purity strongly affects the level of murine protection against tetanus toxin when the toxoid has been applied non-invasively on the skin. (Similar results obtained with injected antigen are not shown).

To substantiate the above mentioned statement, the medium filtrate from a culture of *Clostridium tetani* grown in vitro first was used as an impure antigen. To obtain partially purified antigen, such filtrate was passed through a 10 kDa cut-off membrane and washed thoroughly with phosphate buffer, pH 6.5; in the process, the culture filtrate was concentrated 15 times. Purified toxoid was purchased from Accurate Antibodies, NY, USA.

Swiss albino mice (n = 6) were immunized with identical nominal dose of impure antigen, with partially purified antigen supplemented with monophosphoryl lipid A or with purified antigen with monophosphoryl lipid A added. The antigen was always associated with similar Transfersomes. The composition and the method of preparation for the latter were the same as described with previous examples. The details of immunization schedule, bleeding times and challenge as well as analysis details were also similar to those mentioned before.

The results are given in figure 4. They demonstrate the role antigen purity plays in determining the quality as well as the strength of an immune response against TT. The data shown in figure 4, moreover, indicate that the absorbency of even the specific antibody titre is not a reliable predictor of the therapeutic, that is, of prophylactic effect of an epicutaneous vaccination. This is due to the big differences in specific antibody isotypes which only contain a substantial proportion of Th1-like IgG2b compared to Th2-like IgG1 component if sufficiently pure antigen is used (see also page 12).

Examples 14-15:

Comparison of epicutaneous and subcutaneous administration

Highly deformable vesicles, TransfersomesTM (IDEA):

as described with examples 5-10

Tetanus toxoid dose:

80 μ g TT per epicutaneous immunization

(using 2 mg TT/mL and application area of 2 cm²)

40 μ g TT per subcutaneous injection (using 2 mg TT/mL)

Using the same experimental procedures as described with examples 1-4, as appropriate, the antibody-specific serum titre, the level of animal protection against tetanus toxin and relative occurrence of different specific antibody isotypes was determined.

The results are given in figure 5. While the immunization dependent increase in serum absorbency is comparable after invasive and non-invasive antigen administration the titre in the latter case is somewhat lower by the factor of 6 after primary immunization and by the factor of 8 after second boost. Likewise, while the TT-specific levels of Th2-indicating IgG1 are similar in both arms of this experiment, the specific readings for other antibody subtypes, especially for IgG2a and at early time points also for IgG2b are by the factor 25 and 3, respectively, higher after antigen injections. However, the likelihood for the test mice to survive a subsequent challenge with a normally lethal dose of injected tetanus toxin is independent of the route of antigen administration, within the framework of this experimental series at least.

response within the scope of experimental set-up. Complete animal protection against a normally lethal challenge with 50 LD₅₀ was achieved in this test series with the higher TT dose in combination with LA only.

It was further observed that Th1-cytokine IgG2b was higher with LA groups, compared with the groups that received no LA. This difference was more pronounced for low doses, by the factor of 4, than for high doses, where only an enhancement by the factor of 2 was observed. Th2-cytokine IgG1 was present predominantly, except in the low dose with LA group in which IgG2b contributed comparably.

Examples 22-23:

High molecular weight adjuvant. IL-12 cytokine effect

Highly deformable vesicles, Transfersomes™ (IDEA):

as described with examples 5-10, plus

0.01 mg IL-12 per mL immunogen suspension

Tetanus toxoid, 2 mg/mL, corresponding to 80 µg TT per mouse/ immunization
(partially purified as described with examples 9-11)

Application area: 2 cm² on the upper dorsum of Swiss albino mice.

To study the effect of cytokines on results of non-invasive, epicutaneous vaccination with tetanus toxoid, a combination of monophosphoryl lipid A with 0.4 µg IL-12 per mouse was used. 80 µg of IL-12 was administered per mouse in association with Transfersomes loaded with tetanus toxoid and monophosphoryl lipid A. The details of immunization schedule, bleeding intervals, or the final challenge with the tetanus toxin were the same as mentioned above.

The results of experimental series are illustrated in figure 8. The corroborate the conclusion that the presence of pro Th2 cytokines in the skin during the course of immunopresentation following an epicutaneous TT administration positively affects the outcome of vaccination. This is seen in serum absorbency, the specific antibody titre as well as in the test animal survival probability.

The effect discussed with examples 22-23 was verified by incorporating cytokines other than IL-12 into immunogen formulation. The results are shown in figure 9.

Examples 24-25:

High molecular weight adjuvant (IFN- γ and GM-CSF + IL-4) effect

Highly deformable vesicles, Transfersomes™ (IDEA):

as described with examples 5-8, plus

0.05 mg IFN- γ and 0.004 mg GM-CSF and 0.004 mg IL-4 per mL

immunogen suspension

Tetanus toxoid. 2 mg/mL, corresponding to 80 μ g TT per mouse/ immunization

(impure)

Application area: 2 cm² on the upper dorsum of Swiss albino mice.

The effect discussed with examples 22-23 was confirmed also with a blend of different cytokines. The results are shown in figure 10.

Examples 28-29:

Booster effect (maturation of immune response)

In most of previous examples, a consistent pattern was observed whenever the absorbency was measured during the time course of immunization. The immune response increased with each boost, compared to the response obtained after primary immunization (see Figures 3, 4, 5, 6, 7, 8). The primary response was characterized by predominance of IgM, followed by gradual appearance of IgG after the first boost and by the appearance of even greater amounts of IgG after the second boost, with a concurrent disappearance of IgM. This typical pattern of isotype signifies affinity maturation in the immune response. During the process, the average affinity of a mixture of specific antibodies increases with repeated immunizations.

Results of various epicutaneous vaccination experiments suggest that it may be advantageous to combine an invasive priming vaccination with non-invasive secondary (boost) immunization.

Examples 30-72:Release of cytokines from the skin in vitro by Transfersomes

Highly deformable vesicles (Transfersomes type C):

87.4 mg phosphatidylcholine from soy bean (SPC)

12.6 mg sodium cholate (NaChol)

0.9 mL phosphate buffer, 50 mM, pH 7.3

2.5 μ L thereof

Highly deformable vesicles (Transfersomes type T):

50 mg phosphatidylcholine from soy bean (SPC)

50 mg polysorbate (Tween 80)

0.9 mL phosphate buffer, 50 mM, pH 6.5

2.5 μ L thereof

Positive control A:

2.5 μ L 5% sodium dodecylsulphate (SDS)

Positive control B:

100 μ L lipopolysaccharide (LPS; 10^5 U/mL)

Negative control:

2.5 μ L of phosphate buffered saline (PBS)

All products were tested undiluted.

Cell type: Normal human keratinocytes, forming a pluri-stratified epithelium with a compact stratum corneum were used; histology revealed strong resemblance with human epidermis in vivo.

Method: Keratinocytes were inoculated on polycarbonate filter inserts of 0.63 cm² in chemically defined, supplemented medium, and cultured for 17 days at the air-liquid interface.

Test measurements: given amount of each tested product was deposited with a micropipette and spread evenly over the surface of the stratum corneum of eight reconstituted epidermis using a small sterile device. The cultures were incubated at 37°C, 5% CO₂ for 24 hours. Quadruplicate cultures (except for the LPS treated cells which were incubated in duplicate) were washed with 0.5 mL of PBS and incubated on 300 μ L of 0.5 mg/mL MTT for 3 hours at 37°C, 5% CO₂.

The release of inflammatory mediators (IL1 α , IL2, IL4, IL8, IL10, IFN- γ , and TNF- α) in the medium underlying the tissues was quantified using ELISA kits (R&D systems UK; Quantikine), specific for each type of immuno-modulator to be measured.

	IL1- α (pg/mL) Mean +/- SD	IL8 (pg/mL) Mean +/- SD	TNF- α (pg/mL)
Negative control (PBS, n=2)	5.1 +/- 0.5	< 31	not detectable
Positive control A (SDS 5 %, n=2)	314.2 +/- 6.1	147.5 +/- 32	not detectable
Positive control B (LPS, n=1)	32.0	5161	113.4
Transfersomes C (02-05, n=2)	12.3 +/- 0.9	68.3 +/- 16.8	not detectable
Transfersomes T (TT0009/175, n=2)	11.7 +/- 1.2	50.8 +/- 14.0	not detectable
Transfersomes O (TT0017/15, n=2)	185.5 +/- 170.1	58.4 +/- 27.0	not detectable

The relatively big standard deviation observed with Transfersomes O can be explained by the fact that the product was difficult to spread uniformly onto the stratum corneum of the reconstructed epidermis.

TNF- α level was increased to the level of 113.43 pg/mL when the cells were in contact with the positive controls containing LPS, which is an established immunoadjuvant.

IL8 concentration after cells incubation with Transfersomes exceeded the lower limit of detection by just the factor of 2, which in one case is not and in the other is barely significant at 95% confidence level, but in either situation is negligible compared to the increase observed with the positive control containing the immunoadjuvant LPS, which gave a 167x higher value.

Non-specific irritant, SDS, released a great quantity of IL-1 α from the skin cells into the bathing medium in vitro. The possibility exists, that an amount of comparable quantity was released from the cells incubated with Transfersomes O, comprising the potentially irritating oleic acid at a high concentration, but firm conclusion is prevented by the great standard deviation in the results obtained with the latter test system.

IL-1 α concentration for the other tested Transfersomes of type A and type B changed to approximately 2 times the background level. This difference is statistically significant, compared to negative controls, but practically negligible, taken that the

increase observed with the positive control containing LPS was more than 60 times higher.

IFN- γ , IL-2, IL4 or IL10 was not elevated to a measurable level, suggesting a lack of release of these cytokines, under any other test condition.

Taken together the above mentioned findings suggest that Transfersomes do not release cytokines or induce the generation of such molecules from the skin cells. This explains the need for using immunoadjuvants/modulators when antigens or allergens are to be delivered across the skin with such carriers and elicit a therapeutic or prophylactic immune response.

Examples 73-82:

Bacterial wall component, cholera toxin, as specific immuno-adjuvant:

Highly deformable vesicles, Transfersomes™:

86.3 mg phosphatidylcholine from soy bean (SPC)

13.7 mg sodium cholate (NaChol)

0.9 mL phosphate buffer, 10 mM, pH 6.5

Cholera toxin (CT; Sigma, Neu-Ulm), 10 μ g/immunization plus, if required,

Tetanus toxoid (TT, pure; Accurate Antibodies) 2 mg/mL,

Volume doses corresponding to 0 μ g TT/mouse/immunization (negative control), 1 μ g TT/mouse, 5 μ g TT/mouse, 10 μ g TT/ mouse, 20 μ g TT/ mouse, 40 μ g TT/mouse (in the case of CT usage) and 80 μ g TT/mouse (without CT) were used epicutaneously over an area of up to 2 cm² on the upper dorsum of 4-6 Swiss albino mice; 20 μ g TT/ mouse/immunization were injected subcutaneously at the corresponding site in the positive control group. Unimmunized mice were used as another negative control.

The protective effect of epicutaneous antigen administration was excellent when cholera toxin was included into the test formulation in combination with the tetanus toxoid. Formulation without this immunoadjuvant yielded inferior protection, as demonstrated by the fact that 1 animal out of 4 (25%) was paralytic after the challenge with tetanus toxin.

The results shown in figure 10 reveal that the antigen doses in excess of 20 μg /immunization ensured complete protection, which was not the case with the other tested adjuvants or adjuvant treatments (see previous examples). Lower dosage of antigen gave qualitatively similar effect but was insufficient to guarantee the survival of all test mice, except in the test group which received 5 μg TT/immunization. (This implies that TT doses between 1 μg /immunization and 15 μg /immunization belong to the transition region.) Other doses of cholera toxin might be equally or even more beneficial, however.

Examples 83-85:

Heat labile toxin (HLT) from E.coli has as an immuno-adjuvant

Highly deformable vesicles, Transfersomes™:

86.3 mg phosphatidylcholine from soy bean (SPC)

13.7 mg sodium cholate (NaChol)

0.9 mL phosphate buffer, 10 mM, pH 6.5

Heat Labile Toxin (HLT, SIGMA, Neu-Ulm), 1mg/mL plus, if required

Tetanus Toxoid (TT, pure, Accurate Antibodies) 2 mg/mL

Various relative adjuvant/antigen concentrations were used to test the effect of one of most commonly used ADP ribosylating enzymes for transcutaneous vaccination. Transfersomes were prepared as described in previous examples. Other experimental details (animal housing; antigen administration; titre determination) are described hereinabove.

Volume doses corresponding to 20 μg TT / mouse / e.c. immunization and 1 to 2 μg HLT / mouse / e.c. immunization and a positive control with 0.5 μg TT for s.c. injection were used to immunize Swiss albino mice. The results are illustrated in figure 11.

Anti-TT titres were found to increase when HLT was used as an adjuvant compared to an adjuvant free s.c. injection of antigen or compared to an adjuvant free administration of TT in transfersomes on the skin. Humoral response and the protection against a normally lethal challenge with Tetanus Toxin were found to be dose dependent, with the higher anti-TT titres and improved survival for the higher dose of HLT. The measured results (data not shown) suggest that useful HLT dosage

starts in the range 100 ng/dose while the highest practically useful dose under experimental conditions used in this test series is approx. 100-times greater. This reveals that the upper limit for adjuvant dose that should be used in conjunction with epicutaneously administered Transfersomes is of comparable order of magnitude as, and at most should be 10x higher than, that employed in conventional invasive (s.c.) immunizations; the lower practical limit, in our opinion, is 1-2 orders of magnitude smaller than the typical s.c. dose. It is proposed that similar relationships will also hold for other related immuno-adjuvants.

Examples 86-87:

Local pre-treatment with histamine

Highly deformable vesicles, Transfersomes™:

86.3 mg phosphatidylcholine from soy bean (SPC)

13.7 mg sodium cholate (NaChol)

0.9 mL phosphate buffer, 10 mM, pH 6.5

Tetanus Toxoid (TT, pure, Accurate Antibodies) 2 mg/mL

1 mg/ml histamine solution in phosphate buffer, 10 mM, pH 6.5

In order to test the effect of alternative means for immune-response modulation, mice were injected with 10 μ g histamine in solution at the site of e.c immunization immediately or 15 minute prior to the antigen administration on the skin in Transfersomes. The expectation was that this would induce cytokine release from the skin, and/or have some other positive effect on the outcome of non-invasive transcutaneous immunization. In order to test this hypothesis, Transfersomes were prepared as described in the previously described examples and were carried out with Swiss albino mice as described herein above (except the pre-treatment with histamine).

Figure 12 confirms the above mentioned working hypothesis. The results reveal good humoral response and the requirement for a time period between histamine injection and e.c. immunization to achieve reasonable protection against a challenge with Tetanus Toxin. Comparison with the results measured with TT in Transfersomes on

the skin without an adjuvant thus shows that histamine injection is helpful for boosting animal immune response.

Examples 88-89:

Different administration routes for primary and boost immunization

Highly deformable vesicles, Transfersomes™:

- 86.3 mg phosphatidylcholine from soy bean (SPC)
- 13.7 mg sodium cholate (NaChol)
- 0.9 mL phosphate buffer, 10 mM, pH 6.5
- Cholera Toxin (CT, SIGMA, Neu-Ulm), 0-1 μ g/immunisation
- Tetanus Toxoid (TT, pure, Accurate Antibodies) 2 mg/mL

A detailed description of vesicle preparation and animal experimentation can be found in previous examples hereinabove.

Volume doses corresponding to 20 μ g TT alone or to 0.5 μ g TT plus 1 μ g CT were injected s.c. for primary immunization but were applied epicutaneously for booster immunization. This combination increased the efficacy of vaccination substantially, as implied earlier.

Anti-TT titres were high enough to yield 100% protection against a challenge with Tetanus Toxin (Figure 13). Comparison with the data from previous examples carried out with adjuvant free TT-Tfs on skin clearly demonstrate the usefulness of combined s.c./e.c. vaccination protocol.

The data therefore suggest that antigen-loaded Transfersome™ applied on the skin provide an attractive complement to invasive antigen administration, which should be particularly important for the purpose of booster immunization.

Example 90:

Bi-valent vaccination with Tetanus Toxoid and Cholera Toxin as antigens

Highly deformable vesicles, Transfersomes™:

- 86.3 mg phosphatidylcholine from soy bean (SPC)
- 13.7 mg sodium cholate (NaChol)

50

0.9 mL phosphate buffer, 10 mM, pH 6.5

Cholera Toxin (CT, SIGMA, Neu-Ulm), 10 μ g/immunization

Tetanus Toxoid (TT, pure, Accurate Antibodies) 2 mg/mL

Cholera Toxin not only works as an adjuvant to improve anti-TT response (see previous examples), but is also an antigen by itself. The adjuvant, consequently, can play the role of a secondary antigen, when used at certain concentration. The adjuvancy and immunogenicity are not linearly correlated, however, which offers interesting possibilities for optimizing the outcome of vaccination outcome (also with regard to side effects and allergy induction.) Working with CT in Transfersomes tested in mice, we found out that CT doses between less than 50 ng and at least 10 μ g per application are useful for the purposes described in this application.

Anti-cholera toxin antibodies in the mice that were treated with different volumes of test formulation on the skin (corresponding to 10 μ g TT and 10 μ g CT) are indicative of CT antigenicity. This corroborates the potential of ultradeformable vesicles for making at least bi-valent vaccines based on Transfersomes™ containing more than one antigen.

Figure 14 shows TT and CT titres measured with the mice that were previously immunized with TT and CT in the same carrier.

Luger, T. A., Schwarz, T. The role of cytokines and neuro-endocrine hormones in cutaneous immunity and inflammation. *Allergy* (1995) 50: 292-302.

Wang, L.-F., Lin, J.-Y., Hsieh, K.-H., Lin, R.-H. Epicutaneous exposure of protein antigen induces a predominant Th2-like response with IgE production in mice. *J. Immunol.* (1996) 156: 4079-4082.

CLAIMS

1. A transdermal vaccine comprising
 - (a) a transdermal carrier which is a penetrant, suspended or dispersed in an aqueous solvent, in the form of a minute fluid droplet surrounded by a membrane-like coating of one or several layers of at least two different substances or two different forms of a substance with the tendency to aggregate, said substances or forms of a substance differing by at least the factor of 10 in solubility in a preferably aqueous, liquid medium, such that the average diameter of homo-aggregates of the more soluble substance or form of the substance or the average diameter of the hetero-aggregates consisting of both said substances or forms of said substance is smaller than the average diameter of homo-aggregates of the less soluble substance or form of the substance, and/or wherein the more soluble component tends to solubilise the penetrating droplet and wherein the content of such component amounts to up to 99 mol-% of the concentration required to solubilise the droplet or else corresponds to up to 99 mol-% of the saturating concentration in the un-solubilised droplet, whichever is higher, and/or wherein the elastic deformation energy of the droplet surrounding the membrane-like coating is at least 5x lower, more preferably is at least 10x lower and ideally is more than 10x lower than that of the red blood cells or of the phospholipid bilayers with fluid aliphatic chains;
 - (b) a compound which specifically releases or specifically induces cytokine or anti-cytokine activity or exerts such an activity itself; and
 - (c) an antigen, an allergen, a mixture of antigens and/or a mixture of allergens.
2. The vaccine according to claim 1, wherein the compound displaying or inducing cytokine or anti-cytokine activity and the antigen are associated with the penetrant.
3. The vaccine according to any one of claims 1 or 2, wherein the less soluble self-aggregating molecule is a polar lipid and the more soluble component is a

surfactant or a surfactant-like molecule or else such form of polar lipid which is sufficiently soluble for the purpose of this invention.

4. The vaccine according to any one of claims 1 to 3, wherein the average diameter of the penetrant is between 30 nm and 500 nm, preferably between 40 nm and 250 nm, even more preferably between 50 nm and 200 nm and particularly preferably between 60 nm and 150 nm.
5. The vaccine according to any one of claims 1 to 4, wherein the total weight of droplets in the formulation for the use on human or animal skin is 0.01 weight-% (w-%) to 40 weight-% of total mass, in particular between 0.1 w-% and 30 w-%, and most preferably between 5 w-% and 20 w-%.
6. The vaccine according to any one of claims 1 to 5, wherein total antigen concentration is between 0.001 and 40 w-% of the total penetrant mass, in particular between 0.01 w-% and 30 w-%, even better between 0.1 w-% and 20 w-% and most preferably between 0.5 w-% and 10 w-%.
7. The vaccine according to any one of claims 1 to 6 further comprising
 - (da) a low molecular weight chemical irritant; and/or
 - (db) an extract or a compound from a pathogen or a fragment or a derivative thereof.
8. The vaccine according to any one of claims 1 to 7 wherein the compound exerting cytokine activity is IL-4, IL-3, IL-2, TGF, IL-6, IL-7, TNF, IL-1 α and/or IL-1 β , IL-12, IFN- γ , TNF- β , IL-5 or IL-10 a type I interferon, preferably IFN-alpha or IFN- β .
9. The vaccine according to any one of claims 1 to 8 wherein the compound displaying anti-cytokine activity is an anti-cytokine antibody or the corresponding active fragment, a derivative or an analogue thereof.
10. The vaccine according to any one of claims 1 to 9 wherein the antigen is derived from a pathogen.

11. The vaccine according to claim 10 wherein said pathogen is selected from extracellular bacteria, including pus-forming cocci, such as *Staphylococcus* and *Streptococcus*, gram-negative bacteria, such as *Meningococcus* and *Gonococcus* species, species of *Neisseria*, gram negative bacteria, including enteric organisms such as *E. coli*, *Salmonella*, *Shigella*, *Pseudomonas*, *Diphtheria*, *Bordetella Pertussis*, and gram-positive bacteria (e.g. *Bacillus pestis*, *BCG*), particularly anaerobes, such as the *Clostridium* species (e.g. *Clostridium tetani*, *Clostridium perfringens*, *Clostridium novyi*, *Clostridium septicum*), bacteria and viruses, which survive and replicate within host cells, comprising mycobacteria (e.g. *M. tuberculosis*) and *Listeria monocytogenes*, retro- and adenoviruses, including hepatitis virus, (human) immunodeficiency virus, herpes viruses, small-pox (chicken-pox), influenza, measles, mumps and polio viruses, cytomegalovirus, rhinovirus, etc., and fungi prospering inside host cells, parasites including animal parasites, such as protozoa and helminths, and ectoparasites, such as ticks and mites, or *Brucella* species (e.g. *B. melitensis*, *B. abortus*, *B. suis*, *B. canis*, *B. neotomae*, *B. ovis*), the causative agent for cholera (e.g. *Vibrio cholerae*), *Haemophilus* species like *H. actinomycetemcomitans*, *H. pleuropneumoniae*, as well as pathogens triggering paratyphoid, plague, rabies, tetanus and rubella diseases and pathogens that cause various neoplasiae, auto-immune diseases or are related to other pathological states of the animal or human body which do not necessarily result from pathogen infections.
12. The vaccine according to any one of claims 1 to 11, wherein the allergen is of xenogenic or endogenic origin, derived from a microorganism, an animal or a plant, or belonging to the group of man made and/or irritating inorganic substances, or to such parts or components of the human body which were incorrectly processed by or exposed to the body immune system.
13. The vaccine according to any of claims 1 to 12, wherein the concentration of each compound displaying cytokine activity used is selected to be up to 1000 times higher than the concentration optimum established in the corresponding tests with the antigen dose and immunoadjuvant chosen, performed by injecting

the formulation or performing the tests *in vitro*, and preferably is up to 100x, more often up to 50x and even better up to 20x higher.

14. The vaccine according to any one of claims 7 to 13, wherein the pathogen extract or compound is a lipopolysaccharide, cord-factor (trehalose-dimycolate), muramyl dipeptide, or another (poly)saccharide or (poly)peptide identical to or resembling an immunologically active part of a membrane of a pathogen; an extract of a pathogen, including bacterial exo- and endotoxins, preferably cholera toxin and the heat labile toxin of *E. coli*, an A-chain derivative, a component with an ADP-ribosylating activity, a peptidoglycane, a clostridial toxin, or a purified protein derivative of *M. tuberculosis*, LT-R192G, Fibronectin-binding protein I of *Streptococcus pyogenes*, or outer membrane protein of group B *Neisseria meningitidis* (GBOMP); or bacterial or viral nucleic acids such as oligonucleotides containing unmethylated CpG dinucleotides.
15. The vaccine according to claim 14 wherein said lipopolysaccharide is lipid A or a derivative and modification thereof, such as monophosphoryl lipid A, or its analogue, such as a fatty derivative of saccharose.
16. The vaccine according to any one of claims 7 to 13, wherein the concentration of the pathogen compound derived from a pathogen is between 10x lower and up to 1000x higher than that otherwise used with the corresponding injected formulations employing similar antigen, the epicutaneously administered immunoadjuvant concentration more often differing from the injected immunoadjuvant concentration by the factor between 0.5 and 100, or better, by the factor between 1 and 50, and best between 2 and 25.
17. The vaccine according to any one of claims 7 to 16 wherein said low molecular weight irritant is selected from the classes of allergenic metal ions, acids, bases, irritating fluids, (fatty-) alcohols, (fatty-) amines, (fatty-) ethers, (fatty-) sulphonates, -phosphates, etc., or other suitable solvents or amphiphiles, or from the group of surfactant-like molecules, often with the skin permeation enhancing capability, as well as derivatives or combinations thereof.

18. The vaccine according to any one of claims 7 to 17, wherein the concentration of a low molecular weight irritant is chosen to be by at least the factor of 2, more often by the factor of 5, and even better by the factor of 10 or more, below the concentration which in independent tests on the same or a comparable subject is deemed to be unacceptable owing to the local irritation, as assessed by the methods and standards commonly used to test such an irritant.
19. The vaccine according to any one of claims 7 to 16 wherein the allergen belongs to the class of the inhalation allergens, including but not limited to various pollen, spores, bits of animal hair, skin, feather, natural and synthetic textiles, wheat, (house) dust, including mite; furthermore, food and drug allergens; contact allergens; injection, invasion or depot allergens, such as various (gastrointestine-resident) worms, echinococci, trichines, etc., a part of implantation material.
20. The vaccine according to any one of claims 1 to 19, wherein the applied dose of an antigen differs by the factor of 0.1 to 100 from the dose which otherwise would have to be injected in the process of immunisation, but more often is in the range between 0.5 to 50, even better between 1 and 20 and ideally is less than 10x higher than that used with an injection.
21. The vaccine according to any one of claims 1 to 20, wherein the applied penetrant dose is between 0.1 mg cm^{-2} and 15 mg cm^{-2} , even more often is in the range 0.5 mg cm^{-2} and 10 mg cm^{-2} , and preferably is between 1 mg cm^{-2} and 5 mg cm^{-2} .
22. The vaccine according to any one of claims 1 to 21 wherein said antigen is a pure or purified antigen.
23. A kit comprising, in a bottled or otherwise packaged form, at least one dose of the vaccine according to any one of claims 1 to 22.
24. The kit according to claim 23 further comprising at least one injectable dose of the antigen specified in claim 11 or of the allergen specified in claim 12.

25. A method for generating a protective immuno response on a mammal by vaccinating said mammal with a vaccine according to any one of claims 1 to 22.
26. The method according to claim 25 wherein different treatment areas are selected to control the applied immunogen dose and the outcome of therapeutic vaccination.
27. The method according to claim 25 or 26, wherein a suspension of antigen-free penetrants is loaded with the antigen to be associated therewith during the day prior to an administration, preferably 360 min, more preferably 60 min and even more preferably 30 min before the administration of resulting formulation on the skin.
28. The method according to any one of claims 25 to 27, wherein the vaccine of any one claims 1 to 22 is applied on the skin after pre-treating the organ by an immunoadjuvant manipulation, said manipulation comprising, for example, skin rubbing, pressing, heating, exposing to an electrical or mechanical, e.g. ultrasound field, etc., or injecting a non-immunogenic formulation (like histamine) in the skin, provided that any such treatment releases immunoadjuvant compounds from the skin or other peripheral immuno-active tissues or else reduces the concentration/duration of action of antagonists to the desired vaccination.
29. The method according to any one of claims 25 to 28 wherein immunogen is applied in a non-occlusive patch.
30. The method of any one of claims 25 to 29 characterized in that at least one dose of vaccine is administered.
31. The method according to claim 30 wherein said vaccine is administered as a booster vaccination.
32. The method according to claim 31, wherein the primary immunisation is done

invasively, typically using a subcutaneous injection or some other suitable skin barrier perforating/destructing method, and wherein the at least one subsequent, booster immunisation is done non-invasively.

33. The method according to any one of claims 25 to 32, wherein the vaccine is applied between 2 and 10, preferably between 2 and 7, even more preferably up to 5 and most preferably up to 3 times, when a non-allergenic antigen is used, or such a number of times, in the case of allergens, as is required either to achieve the desired immuno-tolerance, determined according to a suitable assessment method, or else to deem the effort as having failed.
34. The method according to claim 33, wherein the time interval between the subsequent vaccinations is chosen to be between 2 weeks and 5 years, often between 1 month and up to 3 years, more frequently between 2 months and 1.5 years.
35. The method according to any one of claims 25 to 34, wherein the flux of penetrants that carry an immunogen through the various pores in a well-defined barrier is determined as a function of a suitable driving force or a pressure acting across the barrier and the data are then conveniently described by a characteristic curve which, in turn, is employed to optimize the formulation or application further.
36. Use of the transdermal carrier, the compound which specifically releases or specifically induces cytokine or anti-cytokine activity or exerts such an activity, the antigen or allergen, and optionally an extract or a compound from a microorganism or a fragment or a derivative thereof, and/or a low molecular weight chemical irritant as defined in any one of the preceding claims for the preparation of a vaccine for inducing a protective or tolerogenic immune response.

ABSTRACT

The present invention relates to novel vaccines for the non-invasive, transcutaneous administration of antigens associated with ultradeformable carriers, for the purpose of prophylactic or therapeutic vaccination. The vaccines comprise (a) a transdermal carrier which is a penetrant, suspended or dispersed in an aqueous solvent in the form of a minute fluid droplet surrounded by a membrane-like coating of one or several layers of at least two different substances or two different forms of a substance with the tendency to aggregate, said substances or forms of a substance differing by at least the factor of 10 in solubility in a preferably aqueous, liquid medium, such that the average diameter of homo-aggregates of the more soluble substance or form of the substance or the average diameter of the hetero-aggregates consisting of both said substances or forms of said substance is smaller than the average diameter of homo-aggregates of the less soluble substance or form of the substance, and/or wherein the more soluble component tends to solubilise the penetrating droplet and wherein the content of such component amounts to up to 99 mol-% of the concentration required to solubilise the droplet or else corresponds to up to 99 mol-% of the saturating concentration in the un-solubilised droplet, whichever is higher, and/or wherein the elastic deformation energy of the droplet surrounding the membrane-like coating is at least 5x lower, more preferably is at least 10x lower and ideally is more than 10x lower than that of the red blood cells or of the phospholipid bilayers with fluid aliphatic chains, (b) a compound which specifically releases or specifically induces cytokine or anti-cytokine activity or exerts such an activity itself, and (c) an antigen, an allergen, a mixture of antigens and/or a mixture of allergens. The invention further relates to methods for the vaccination of mammals for obtaining a protective or therapeutic immune response.

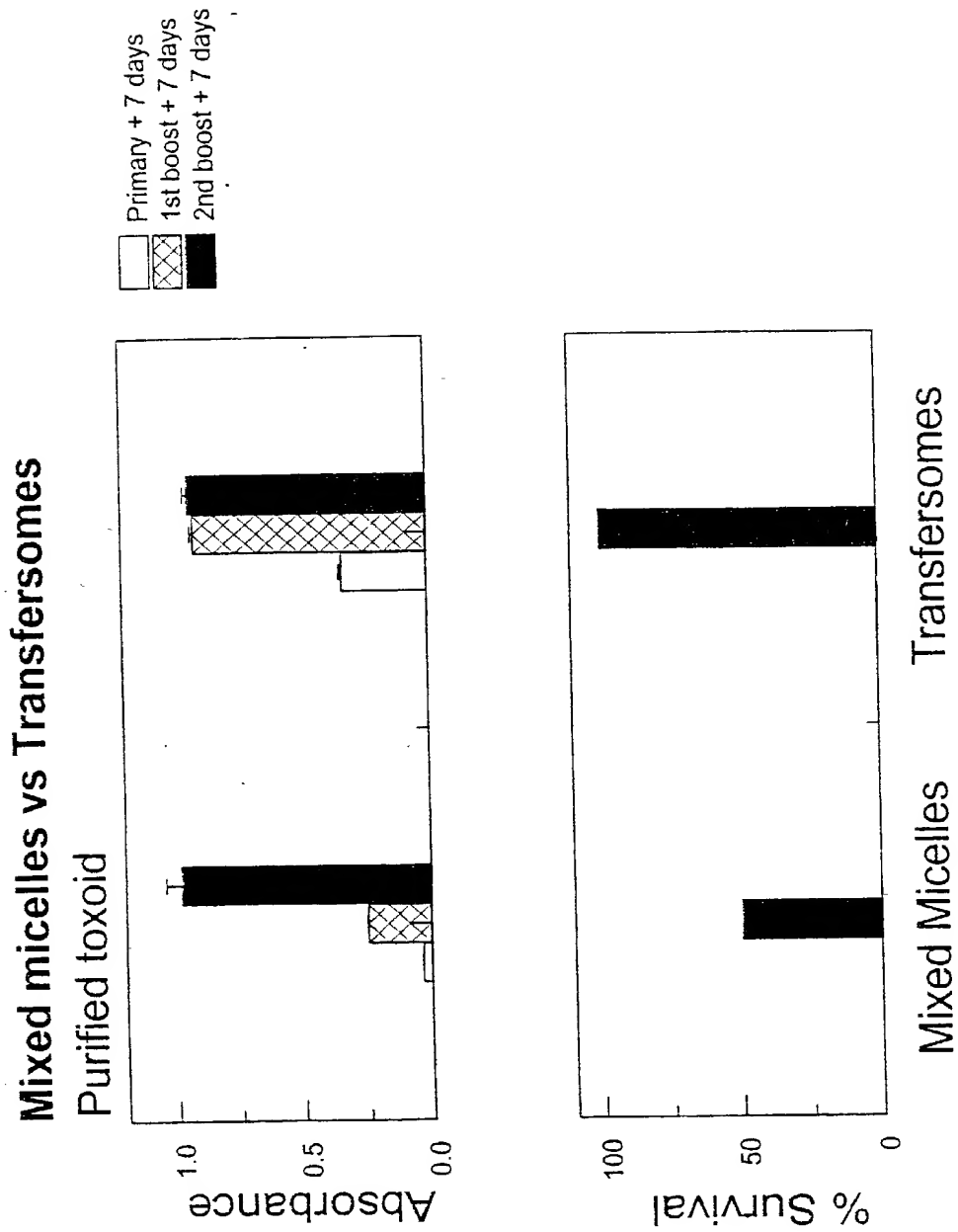


Figure 1

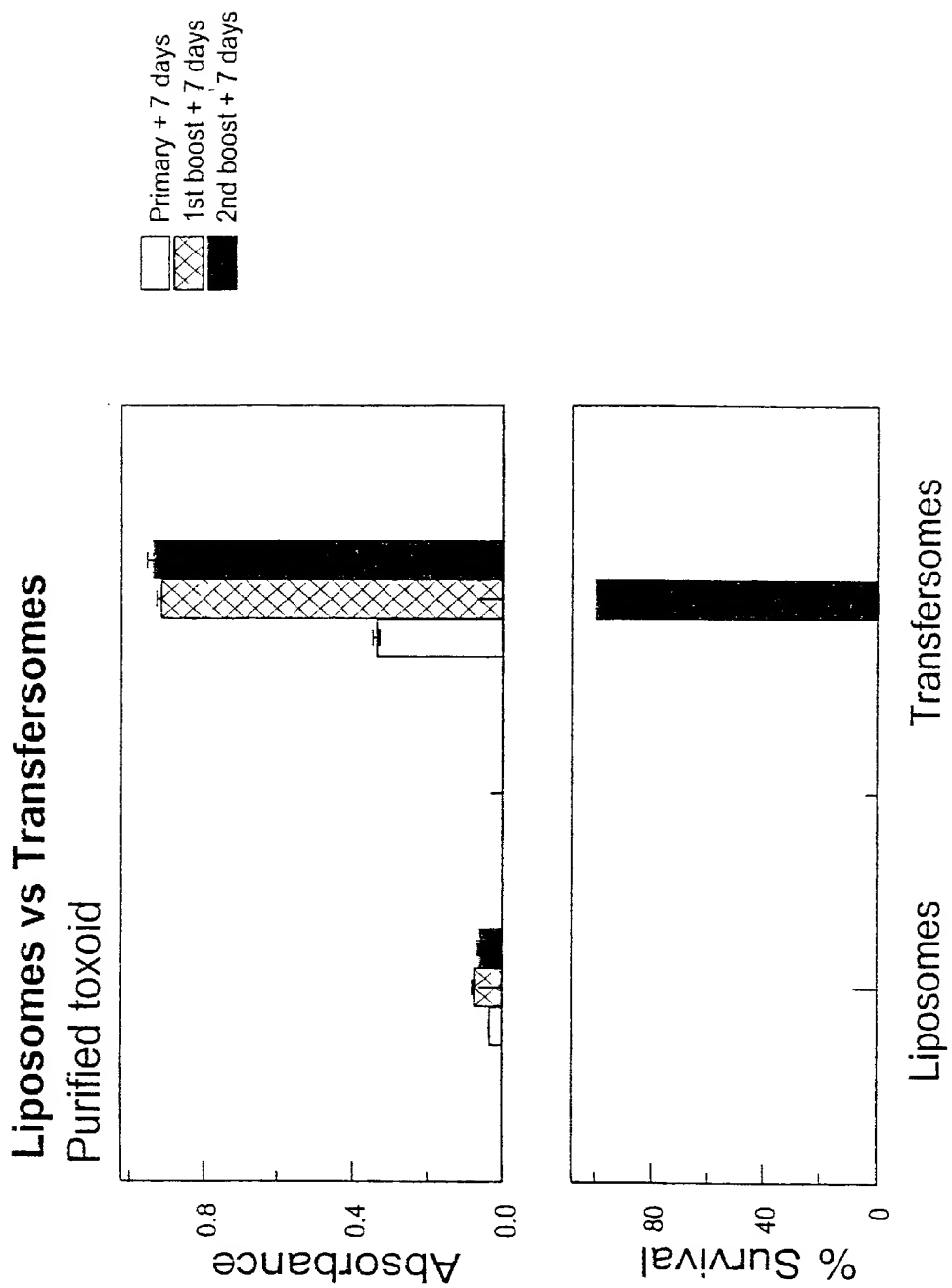


Figure 2

Antigen Dose Effect

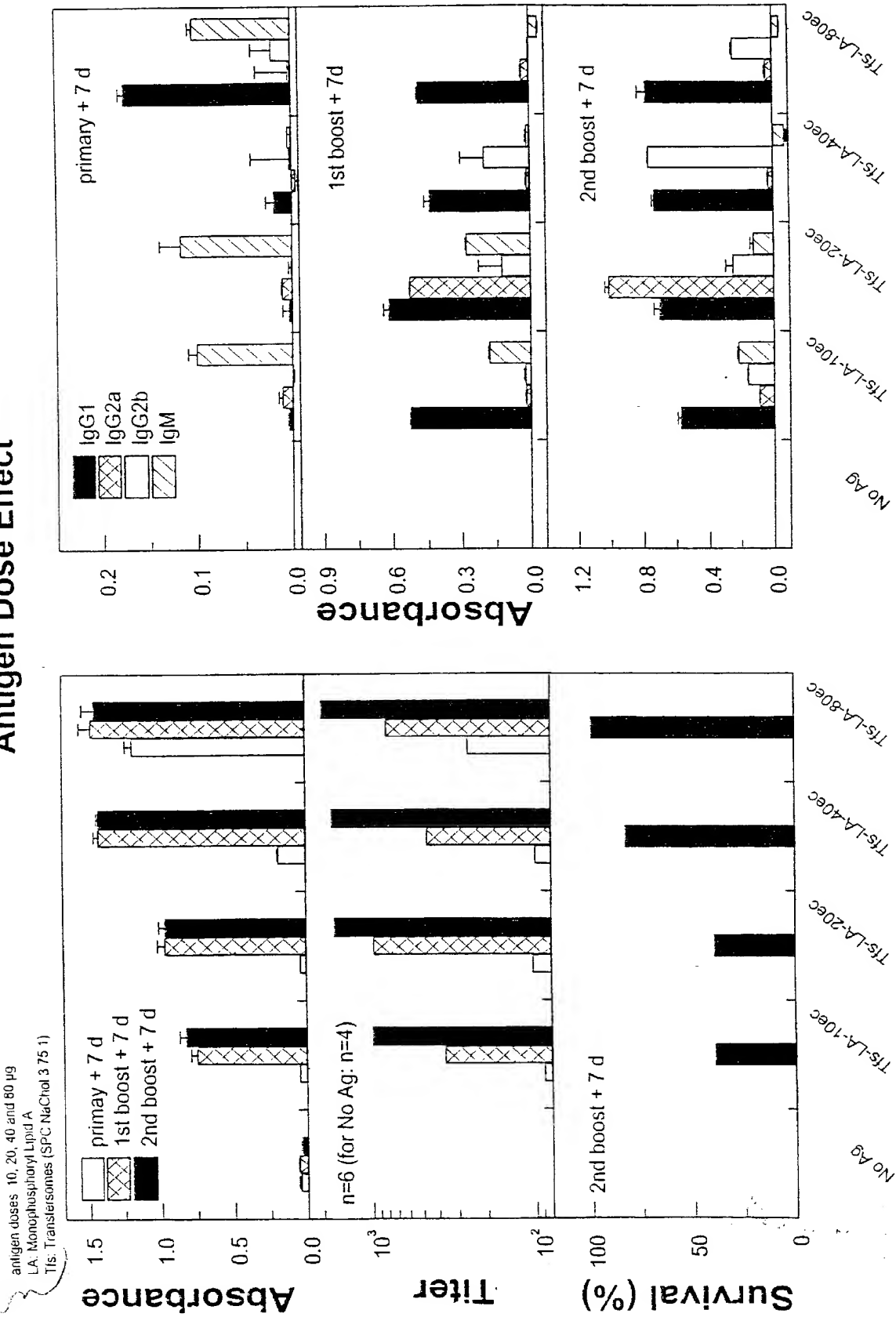


Figure 3

Antigen Purity Effect

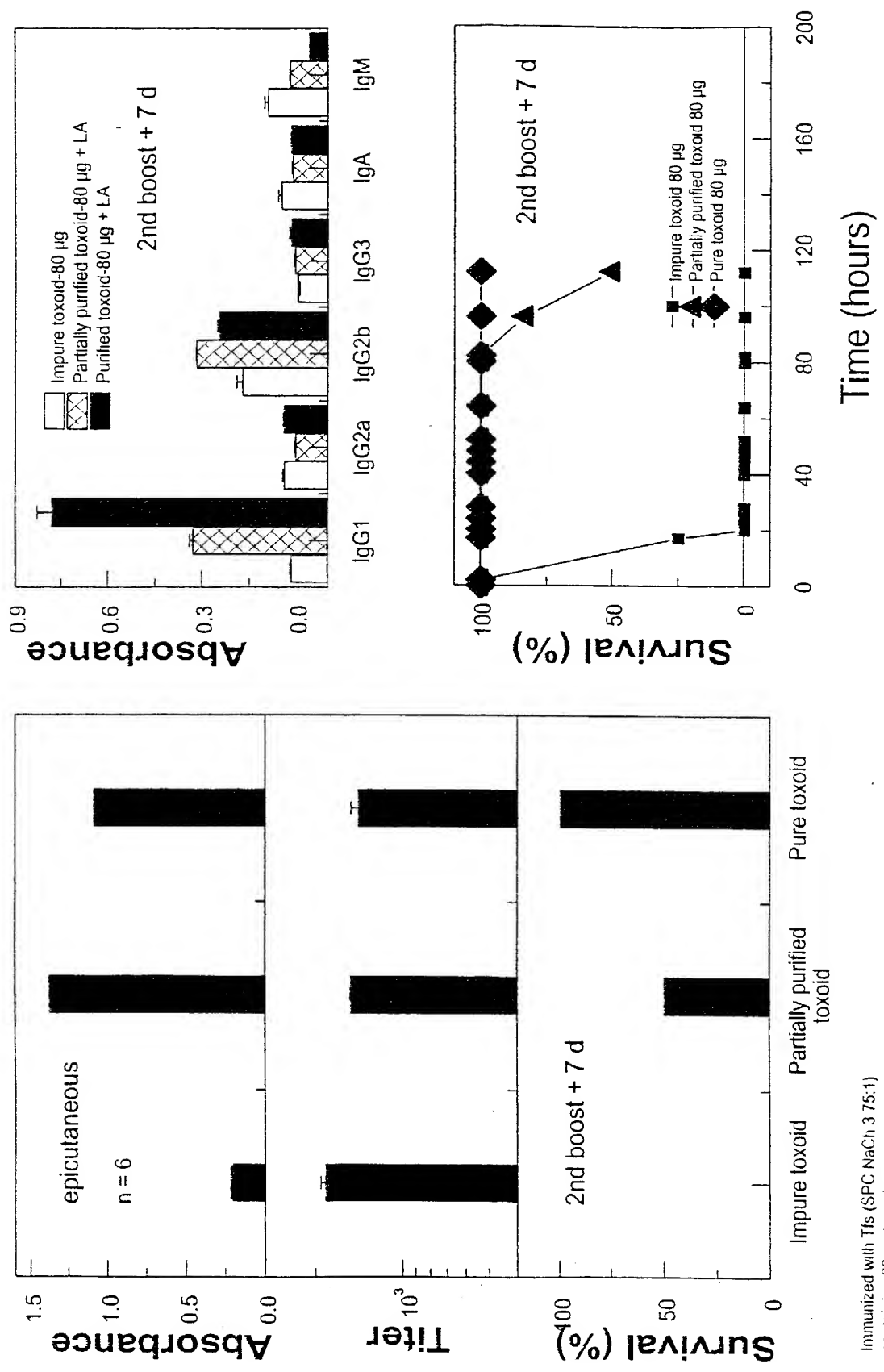


Figure 4

Immunized with Tfs (SPC NaCh 3 75:1)
containing 80 µg toxoid
LA: Monophosphoryl lipid A

Epicutaneous vs. Subcutaneous Immunisation

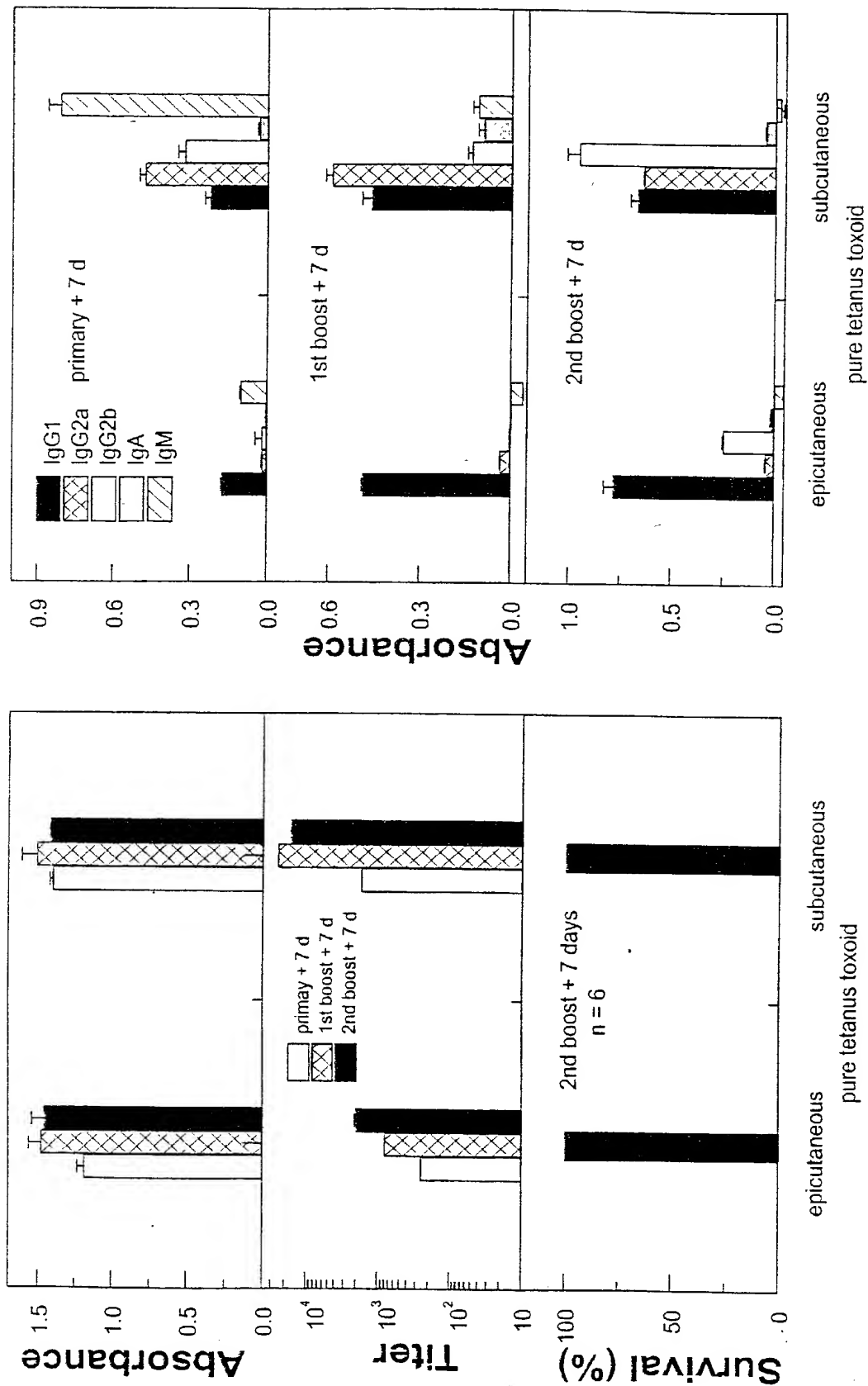


Figure 5

Pre-injection Effect

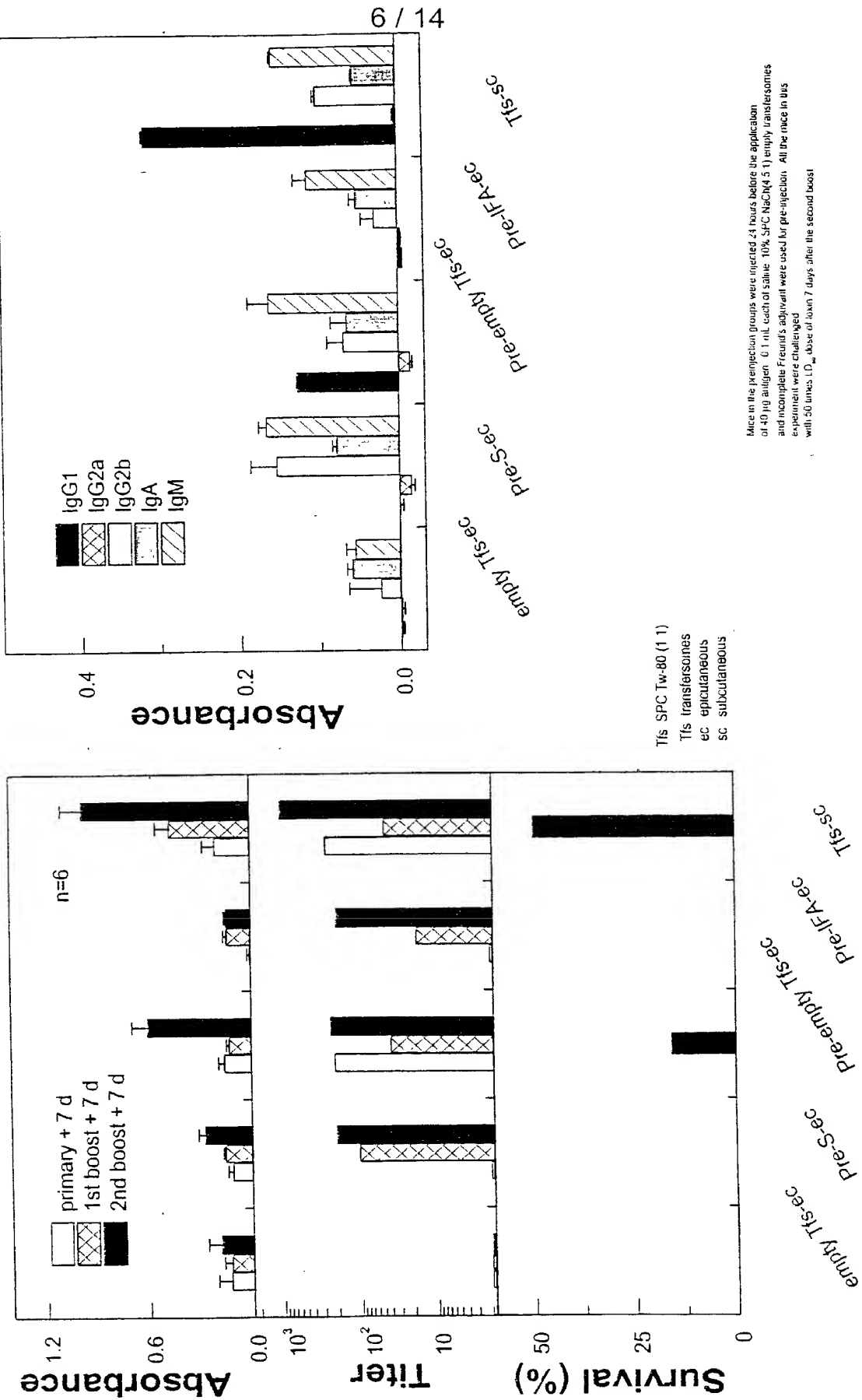


Figure 6

Adjuvant effect: for example, monophosphoryl Lipid A (LA)

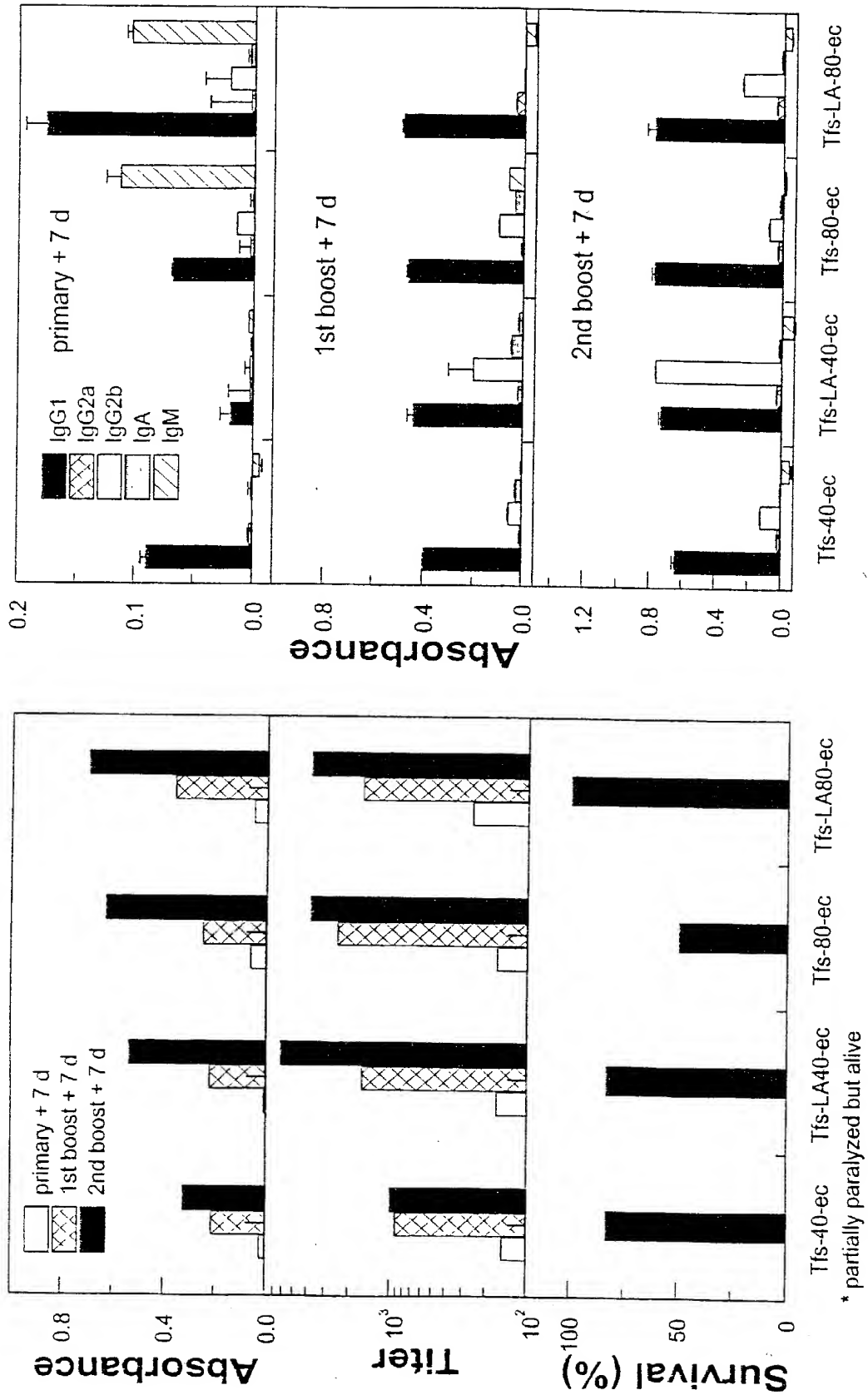


Figure 7

* partially paralyzed but alive

Adjuvant effect: for example cytokine IL-12

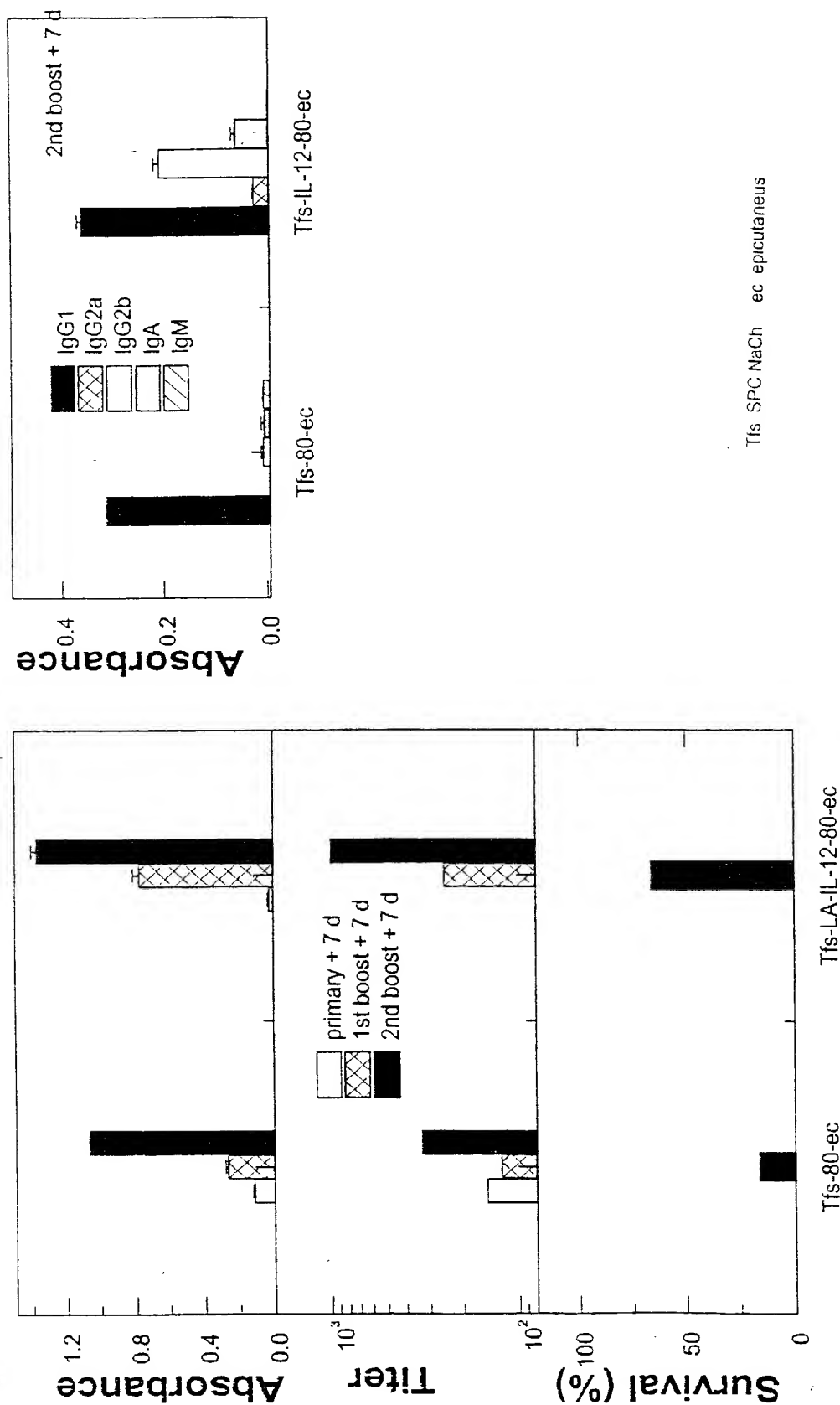
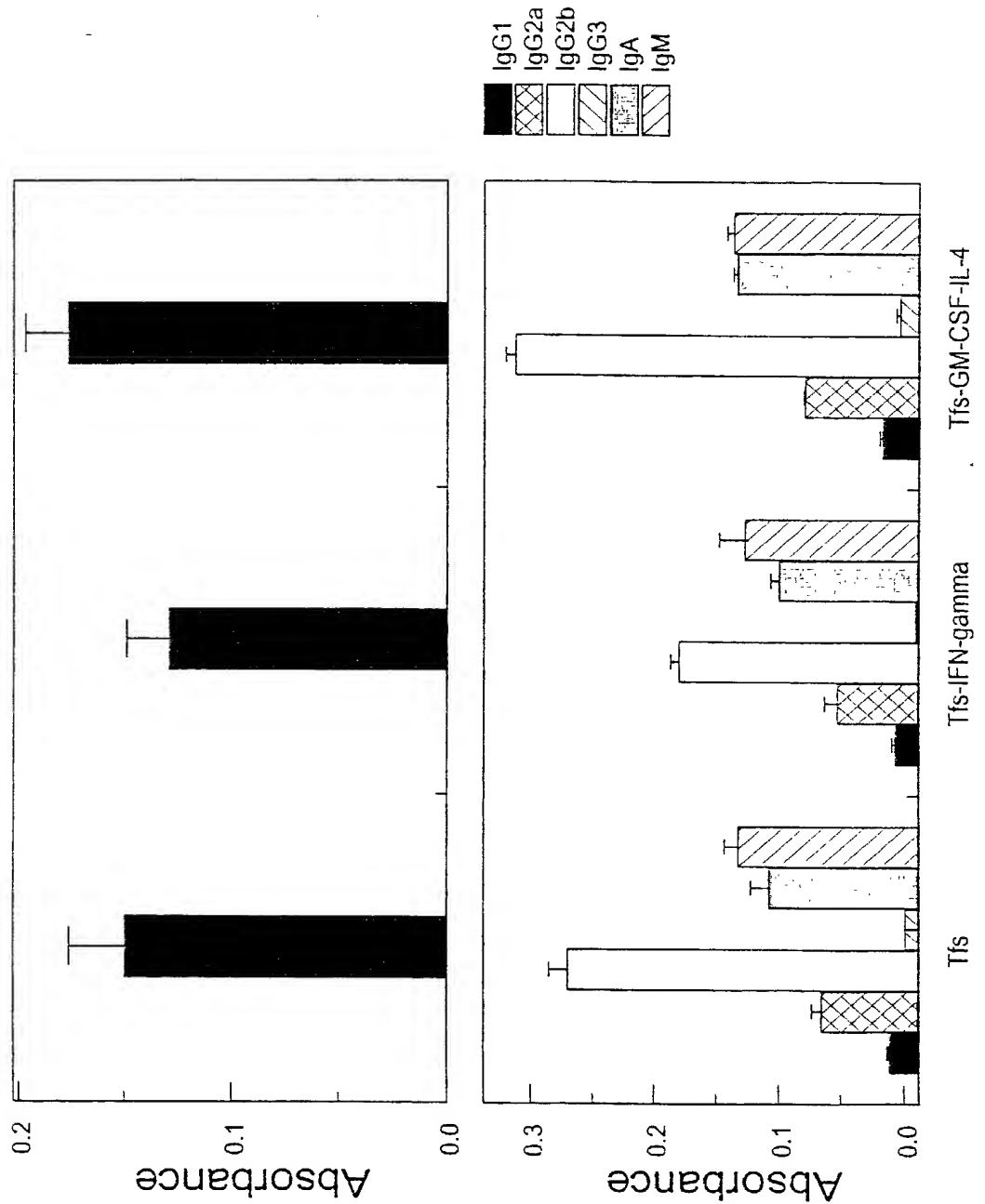


Figure 8

Immunomodulant effect, for example cytokines

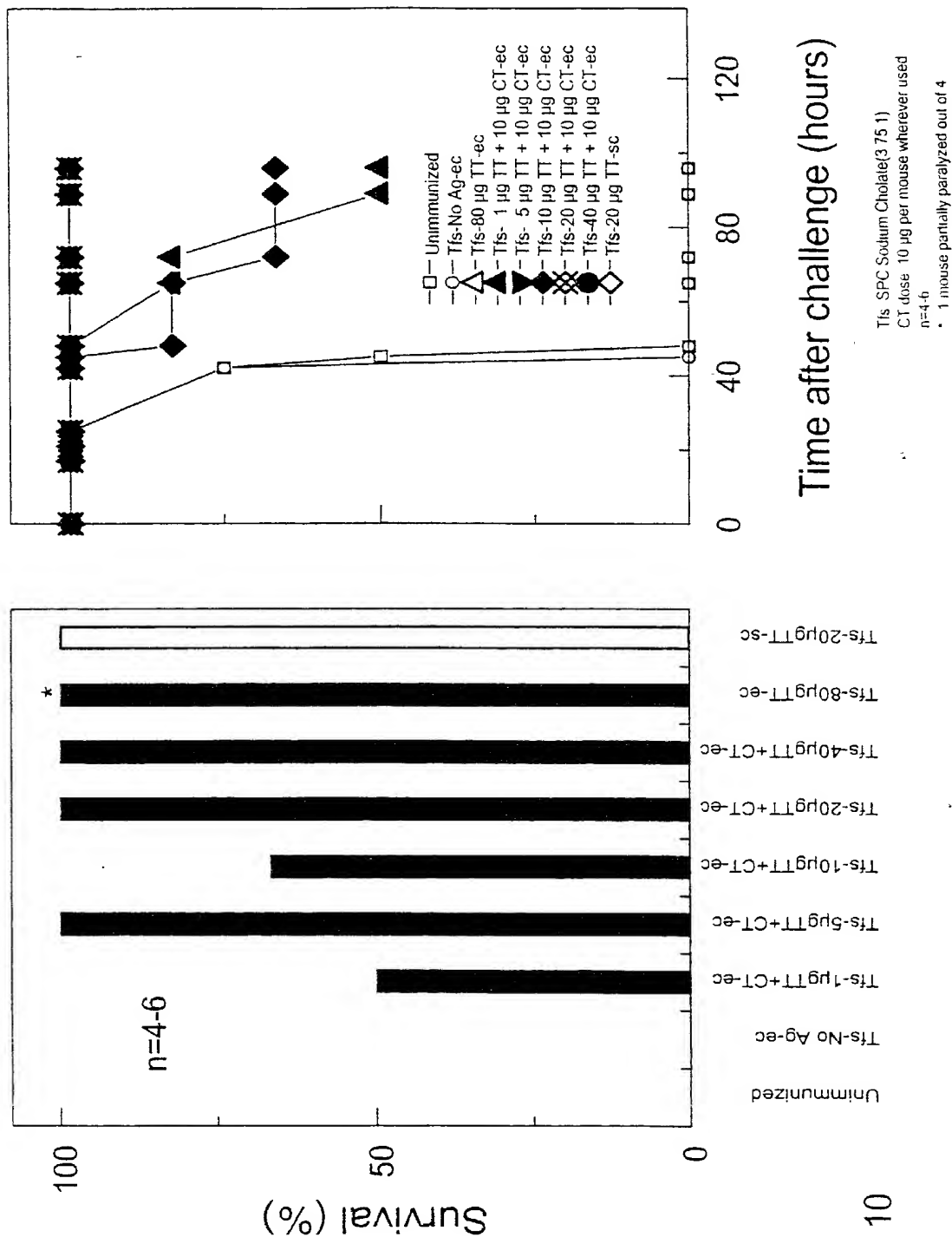
Impure toxoid



Serum was collected for the assay on 7th day after 2nd boost
No protection was observed in any of the groups

Figure 9

Immunoadjuvant effect: for example, cholera toxin (CT)
added to tetanus toxoid (TT, pure) in Transfersomes (Tfs) on the skin



Adjuvant effect: for example Heat Labile Toxin (HLT) from E.coli

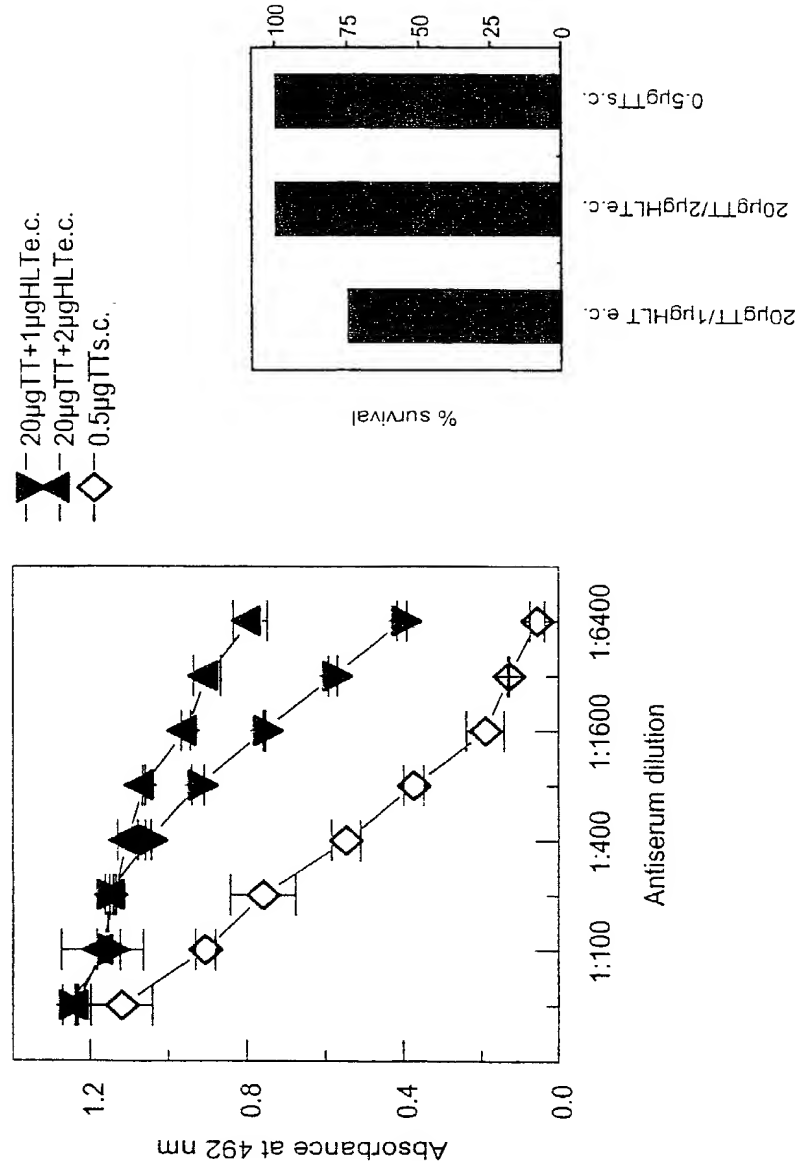
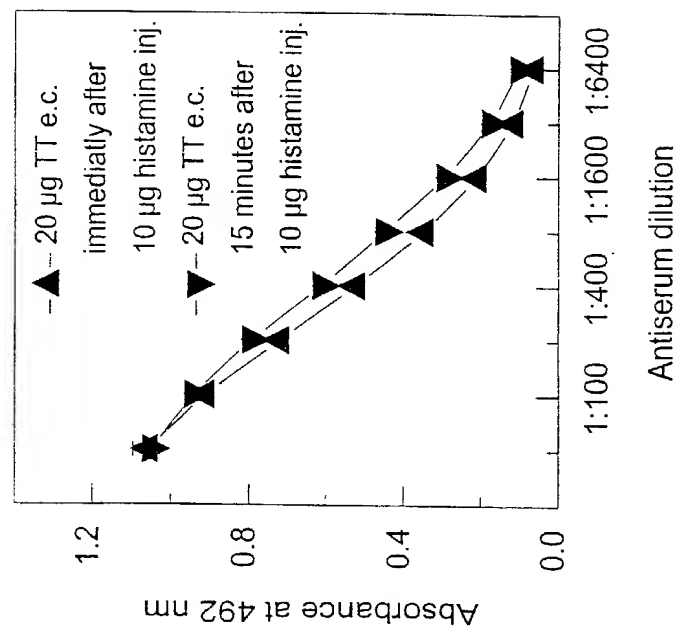


Figure 11

Histamine effect: on anti-tetanus titer and survival after immunization with Transfersomes on the skin



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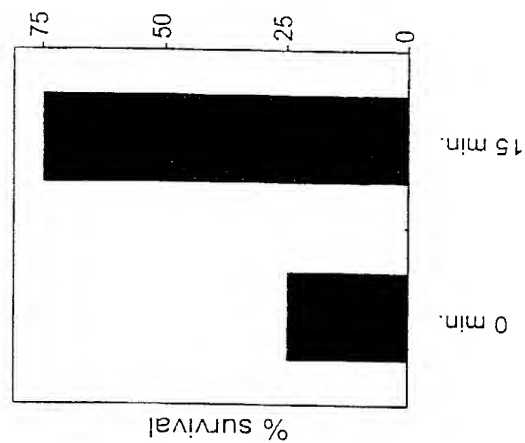


Figure 12

Subcutaneous priming: effect on anti-tetanus titer and survival after epicutaneous boosts

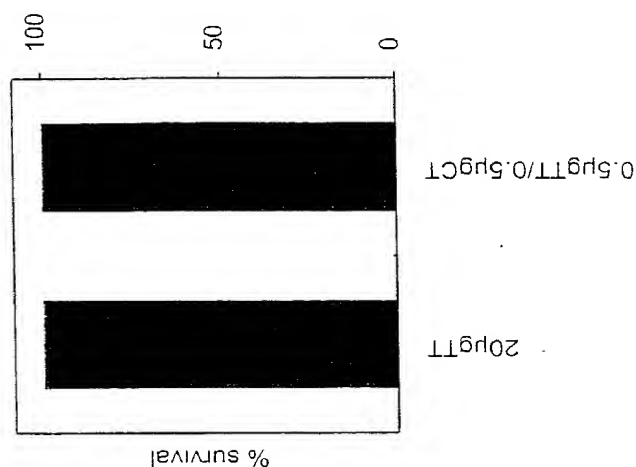
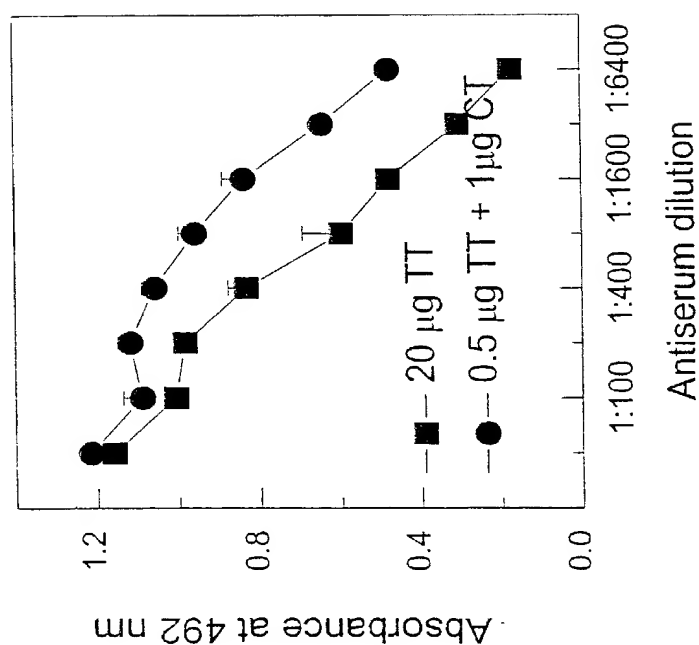


Figure 13

Bi-Valent Vaccines: Anti-Tetanus and anti-Cholera
response to the administration of both antigens together
in Transfersomes on the skin

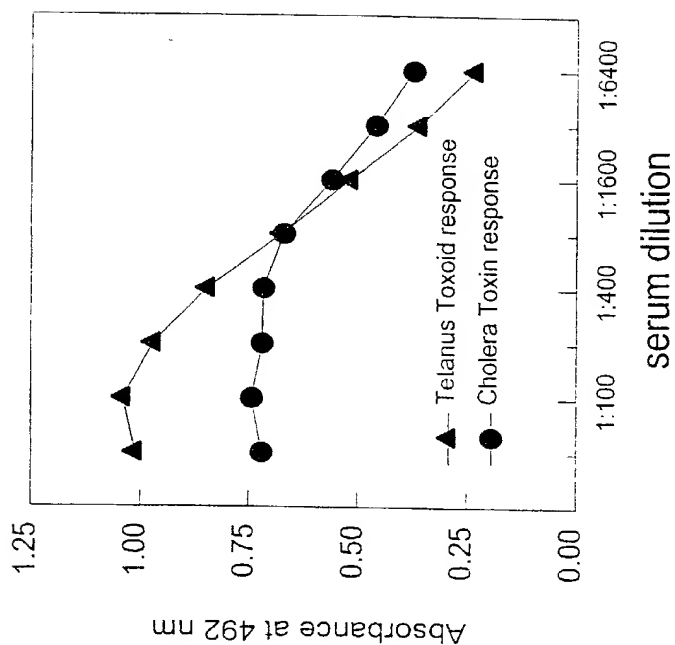


Figure 14